

**ENHANCEMENT OF ADENOVIRAL ONCOLYTIC ACTIVITY IN PROSTATE**  
**CELLS BY MODIFICATION OF THE E1A GENE PRODUCT**

**Related Applications**

This application claims the benefit of U.S. Provisional Application Serial No. 60/401,919, filed August 8, 2002. The entire contents of this application is incorporated herein by this reference.

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**Background of the Invention**

Prostate cancer is the most common cancer, excluding non-melanoma skin cancers, in American men. The American Cancer Society estimates that in the year 2000 approximately 180,400 new cases of prostate cancer were diagnosed in the United States. Prostate cancer is the second leading cause of cancer death in men, exceeded only by lung cancer. Prostate cancer causes about 11 percent of all cancer deaths in men. Furthermore, it is estimated that approximately 5 million men have at this very moment a histological cancer of the prostate, which may or may not ever become clinically evident. The prostate gland is about the size of a walnut and is located anterior to the rectum, adjacent to the bladder and surrounds the proximal part of the urethra. It contains glandular epithelium that produces a portion of the seminal fluid which protects and nourishes sperm cells. Although other cells exist in the prostate, over 99% of prostate cancers develop are adenocarcinomas.

Prostate cancer is a multi-focal disease with clones of androgen-sensitive and androgen-refractory cells (N. Kyprianou and J. Isaacs, Biochem. Biophys. Res. Comm., 165, 73-81, 1989; Kyprianou, N., et al., World J. Urol., 12, 299-303, 1994; M. Tenniswood and H. Michna, Ernst Schering Research Foundation Workshop, 14, Springer-Verlag, Berlin Heidelberg, 1995). The role androgen receptors (AR) may play in prostate cancer is not clear.

Although androgen depletion therapy results in regression of the tumor, it returns as an androgen-refractory cancer. Some androgen-refractory tumors express increased levels of androgen receptors, suggesting that continued proliferation of androgen-refractory prostate cells may be influenced by androgens (Linja, M. J., et al., *Cancer Res.*, 61, 3350-3555, 2001).

5 Paradoxically, increased expression of AR may also be responsible for inhibition of growth and may induce apoptosis (Joly-Pharaboz, M. O., et al., *J. Steroid Biochem. Mol. Biol.*, 55, 67-76, 1995; Joly-Pharaboz, M. O., et al., *J. Steroid Biochem. Mol. Biol.*, 73, 237-249, 2000; Dai, J. L., et al., *Steroids*, 61, 531-539, 1996; Umekita, Y., et al., *Proc. Natl. Acad. Sci., USA*, 93, 11802-11807, 1996; Zhau, H. Y. E., et al., *Proc. Natl. Acad. Sci., USA*, 93, 15152-10 15157, 1996; Heisler L. E., et al., *Mol. Cell. Endocr.*, 126, 59-73, 1997; Shen, R., et al., *Endocrinology*, 141, 1699-1704, 2000).

Patients diagnosed with a clinically localized or "early-stage" prostate cancer may be treated with surgery, radiation, local ablation, or by non-treatment or "watchful waiting." The 15 conventional surgery is a radical prostatectomy, which can be performed through a retropubic approach, a perineal approach, or in the case of laparoscopy through a transperitoneal approach. Surgery is usually reserved for locally confined disease and is usually curative. (Catalona W J et al., "Contemporary results of anatomic radical prostatectomy," *CA Cancer J Clin* 40: 282, 1999). These procedures are invasive, possess significant side effects (urinary 20 incontinence and impotence), and require hospital stays and time out of work. Definitive surgical treatment to extirpate the early stage cancer often involves some type of radical prostatectomy. Modifications of surgical techniques have been developed to preserve potency in patients undergoing radical prostatectomy. (Walsh, P. C. "Anatomic radical prostatectomy: evolution of the surgical technique". *J Urol*, 160: 2418-2424, 1998.). Meticulous surgical 25 technique is vital, however, to minimize the incidence of positive surgical margins and consequent recurrent disease. (Rosen M A et al., "Frequency and location of extracapsular extension and positive surgical margins in radical prostatectomy specimens," *J. Urol.* 148: 331, 1992).

30 Considerable interest has evolved in developing gene therapy vectors as therapeutic agents. Many proposed cancer therapeutic vectors are based on adenovirus. U.S. Pat. Nos. 5,631,236 and 6,096,718 (Baylor College of Medicine) cover a method of causing regression in a solid tumor, using a vector containing an HSV thymidine kinase (tk) gene, followed by administration of a prodrug such as ganciclovir. U.S. Pat. No. 6,096,718 (Baylor College of

Medicine) relates to the use of a replication incompetent adenoviral vector, comprising an HSV tk gene under control of the alpha-lactalbumin promoter. U.S. Pat. Nos. 5,801,029 and 5,846,945 (Onyx Pharmaceuticals) relate to adenovirus in which the E1b gene has been altered so as not to bind and inactivate tumor suppressor p53 or RB proteins expressed by the host. This prevents the virus from inactivating tumor suppression in normal cells, which means the virus cannot replicate. However, the virus will replicate and lyse cells that have shut off p53 or RB expression through oncogenic transformation.

Adenoviruses are non-enveloped, regular icosohedral, double-stranded DNA viruses.

10 The protein coat (capsid) is composed of 252 capsomeres of which 240 are hexons and 12 are pentons. Most of the detailed structural studies of the adenovirus polypeptides have been done for adenovirus types 2 and 5. The viral DNA is 23.85 X10<sup>6</sup> daltons for adenovirus 2 and varies slightly in size depending on serotype. The DNA has inverted terminal repeats and the length of these varies with the serotype.

15 The replicative cycle is divided into early (E) and late (L) phases. The late phase defines the onset of viral DNA replication. Adenovirus structural proteins are generally synthesized during the late phase. Following adenovirus infection, host DNA and protein synthesis is inhibited in cells infected with most serotypes. The adenovirus lytic cycle with adenovirus 2 and adenovirus 5 is very efficient and results in approximately 10,000 virions per infected cell along with the synthesis of excess viral protein and DNA that is not incorporated into the virion. Early adenovirus transcription is a complicated sequence of interrelated biochemical events, but it entails essentially the synthesis of viral RNAs prior to the onset of viral DNA replication.

20 25 The organization of the adenovirus genome is similar in all of the adenovirus groups and specific functions are generally positioned at identical locations for each serotype studied. Early cytoplasmic messenger RNAs are complementary to four defined, noncontiguous regions on the viral DNA. These regions are designated (E1-E4). The early transcripts have been classified into an array of immediate early (E1a), delayed early (E1b, E2a, E2b, E3 and E4), and intermediate (IVa2.1X) regions.

The E1a region is involved in transcriptional transactivation of viral and cellular genes as well as transcriptional repression of other sequences. The E1a gene exerts an

important control function on all of the other early adenovirus messenger RNAs. In normal tissues, in order to transcribe regions E1b, E2a, E2b, E3, or E4 efficiently, active E1a product is required. However, the E1a function may be bypassed. Cells may be manipulated to provide E1a-like functions or may naturally contain such functions. The virus may also be  
5 manipulated to bypass the functions.

The E1b region is required for the normal progression of viral events late in infection. The E1b product acts in the host nucleus. Mutants generated within the E1b sequences exhibit diminished late viral mRNA accumulation as well as impairment in the inhibition of  
10 host cellular transport normally observed late in adenovirus infection (Berkner, K. L., Biotechniques 6:616-629 (1988)). E1b is required for altering functions of the host cell such that processing and transport are shifted in favor of viral late gene products. These products then result in viral packaging and release of virions. E1b produces a 19 kD protein that prevents apoptosis. E1b also produces a 55 kD protein that binds to p53.

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For a complete review on adenoviruses and their replication, see Horwitz, M. S., Virology 2d ed, Fields, B. N., eds., Raven Press Limited, New York (1990), Chapter 60, pp. 1679-1721.

20 Until relatively recently, the virtually exclusive focus in development of adenoviral vectors for gene therapy has been use of adenovirus merely as a vehicle for introducing the gene of interest, not as an effector in itself. Replication of adenovirus had previously been viewed as an undesirable result, largely due to the host immune response. More recently, however, the use of adenovirus vectors as effectors has been described (see e.g., International  
25 Patent Application Nos. PCT/US98/04084, PCT/US98/04080; PCT/US98/04133, PCT/US98/04132, PCT/US98/16312, PCT/US95/00845, PCT/US96/10838, PCT/EP98/07380, U.S. Pat. No. 5,998,205 and U.S. Pat. No. 5,698,443). The use of IRES in vectors have been described. See, for example, International Patent Application No. PCT/US98/03699 and International Patent Application No. PCT/EP98/07380. Adenovirus  
30 E1A and E1B genes are disclosed in Rao et al. (1992, Proc. Natl. Acad. Sci. USA vol. 89: 7742-7746).

Publications describing various aspects of adenovirus biology and/or techniques relating to adenovirus include the following: PCT/US95/14461; Graham and Van de Eb

(1973) Virology 52:456-467; Takiff et al. (1981) Lancet 2(8251):832-834; Berkner and Sharp (1983) Nucleic Acid Research 11 (17);6003-6020; Graham (1984) EMBO J 3:2917-2922; Bett et al. (1993) J. Virology 67:5911-5921; and Bett et al. (1994) Proc. Natl. Acad. Sci. USA 91:8802-8806. These references describe adenoviruses that have been genetically modified to produce replication-defective gene transfer vehicles. In such vehicles, the early adenovirus gene products E1A and E1B are deleted and provided in trans by the packaging cell line 293 developed by Frank Graham (Graham et al. (1987) J. Gen. Biol. 36:59-72 and Graham (1977) J. Genetic Virology 68:937-940). The gene to be transduced is commonly inserted into adenovirus in the deleted E1A and E1B region of the virus genome Bett et al. (1994), supra. Adenovirus vectors as vehicles for efficient transduction of genes have been described by Stratford-Perricaudet (1990) Human Gene Therapy 1:2-256; Rosenfeld (1991) Science 252:431-434; Wang et al. (1991) Adv. Exp. Med. Biol. 309:61-66; Jaffe et al. (1992) Nat Gen. 1:372-378; Quantin et al. (1992) Proc Natl. Acad. Sci. USA 89:2581-2584; Rosenfeld et al. (1992) Cell 68:143-155; Stratford-Perricaudet et al. (1992) J. Clin. Invest. 90:626-630; Le Gal La Salle et al. (1993) Science 259:988-990; Mastrangeli et al. (1993) J. Clin. Invest. 91:225-234; Ragot et al. (1993) Nature 361:647-650; Hayashi et al. (1994) J. Biol. Chem. 269:23872-23875.

There are several other experimental cancer therapies which utilize various aspects of adenovirus or adenovirus vectors. See, U.S. Pat. No. 5,776,743; U.S. Pat. No. 5,846,945; U.S. Pat. No. 5,801,029; PCT/US99/08592; U.S. Pat. No. 5,747,469; PCT/US98/03514; and PCT/US97/22036.

Of particular interest is the development of more specific, targeted forms of prostate cancer therapy. In contrast to conventional cancer therapies, which result in relatively non-specific and often serious toxicity, more specific treatment modalities attempt to inhibit or kill malignant cells selectively while leaving healthy cells intact. There is, therefore a serious need for developing specific, less toxic prostate cancer therapies.

Furthermore, an effective treatment for prostate cancer will kill both androgen-responsive and androgen-refractory cancer cells. Whereas commonly used androgen deprivation therapies induce apoptotic cell death in androgen-sensitive cells (Colombel, M. C., et al., Methods Cell. Biol., 46, 27-34, 1995; Buttyan, R., et al., In: Prostate--Basic and Clinical Aspects., pp 201-218, Naz R K (ed), CRC Press, Boca Raton, 1997; Perlman, H., et

al., Cell Death Differentiation 6, 48-54, 1999; Bruckheimer, E. M., et al., Sem. Oncol., 26, 382-398, 1999), effective chemotherapy for androgen-refractory cancer is not available (Kozlowski, J., et al., Urol. Clin. N. Am., 18, 15-24, 1991; Santen, R. J., J. Clin. Endocrinol. Metab., 75, 685-689, 1992; Kreis, W., Cancer Invest., 13, 296-312, 1995). It would be  
5 advantageous to have means of inducing cell death in all prostate cancer cell types, including both androgen-sensitive and androgen-refractory prostate cancer cells.

All references cited herein are hereby incorporated by reference in their entirety.

10 **Summary of the Invention**

The invention provides compositions and methods for the administration of prostate cell-specific adenoviral vectors to an individual in need thereof, such as, an individual with prostate cancer.

15 Accordingly, in one aspect, the invention provides compositions and methods of suppressing prostate tumor growth in an individual comprising administering to the individual a composition comprising a replication-competent target cell-specific adenoviral vector wherein said vector comprises a DNA sequence encoding a chimera of an adenovirus gene essential for replication, specifically the E1A gene, and the gene encoding the androgen  
20 receptor (or a portion thereof) under transcriptional control of a target cell specific transcriptional regulatory element (TRE), wherein the adenoviral vector is administered in amounts sufficient to suppress prostate tumor growth

Any TRE which directs prostate cell-specific expression can be used in the disclosed  
25 adenovirus vectors. TREs specific for prostate cancer cells include, the rat probasin (PB) TRE; prostate-specific antigen (PSA) TRE; the hK2 TRE, or any prostate specific promoter which contains an androgen responsive element.

In other embodiments, the chimera is comprised of sequences of the E1A gene and  
30 fragments of the androgen receptor gene, such as the transactivation domain and the DNA Binding Domain. In another embodiment, there is a single point mutation the full length amino acid sequence of the Androgen receptor in the E1A/AR chimeric protein. The single point mutation occurs at the codon 685 position, where cysteine is replaced by tyrosine, which results in a chimera activated by both androgens and nonsteroidal anti-androgens.

In still another embodiment, the E1A/Androgen receptor chimera can be inactivated by ansamycins, such as geldanamycin.

Other features and advantages of the invention will be apparent to those skilled in the  
5 art from the following detailed description and claims.

#### Brief Description of the Drawings

Figure 1 A is bar graph representations illustrating E1A inhibition of AR function in LAPC4; in A, E1A expressed either by plasmid in a cotransfection, or by virus in an early  
10 infection response causes a loss of AR inducibility for the target reporter pBK-PSE-PBN-LUC; Viral experiments were performed at varying multiplicities of infection, shown in the grids below the graph; B is bar graph representations illustrating the effect of E1A on expression of an AR dependent promoter construct in an AR negative cell line (PC3); note that the overall activity is diminished compared to LAPC4 and there is no significant effect of  
15 E1A expression either by plasmid or by virus on the target reporter pBK-PSE-PBN-LUC at this concentration of E1A plasmid (0.1 ug); because virus enhances plasmid DNA transfer efficiency, data are plotted as fold induction of RLU relative to the negative control.

Figure 2 is bar graph representations illustrating E1A Mediated Inhibition of Various  
20 Prostate Specific Promoters; three basic prostate specific promoters (rat Probasin, human PSA, and human PSMA) were used in this analysis, as well as chimeric constructs containing the enhancer element from the PSA gene (-5322 to -3738) fused to the different promoters; these promoter constructs were placed upstream of the luciferase reporter gene; all values reported are normalized using a co-transfected Renilla luciferase reporter; all values reported  
25 were performed in the presence of 10 nM R1881; in the appropriately marked lanes, an expression plasmid encoding viral genomic E1A under the control of the CMV promoter was co-transfected at a high level compared to previous experiments (0.5 mcg/well); in A, LNCaP (AR positive) cells demonstrated that the addition of AREs (via the PSE) resulted in augmentation of the PSA promoter and the probasin promoter, but not the PSMA promoter;  
30 overexpression of E1A resulted in marked inhibition of expression of the reporters; in B, PC3 (AR minus) demonstrated less total promoter activity, as expected; the addition of AREs (via the PSE) did not result in significant augmentation of transcriptional activity, as there is no AR; overexpression of E1A, however, did demonstrate a decrease in total promoter activity; these results indicate that high levels of E1A can suppress transcription

from a variety of prostate specific promoters, even without involving the androgen receptor; RLU are expressed as % Renilla control co-transfection expression (i.e., dual luciferase).

5 Figure 3 is bar graph representations illustrating E1A fusions with androgen receptor domains; the various different fusions were tested in 3 different cell lines, in the absence and presence of 10 nM R1881; in A, LNCaP B. PC-3. C. HeLa as can be seen, the LNCaP showed remarkable induction of an AR dependent promoter (pBK-PSE-PBN-Luc) with the E1A-AR fusion; a similar effect was seen with PC-3, but to a slightly lesser degree as there is no endogenous AR to complex with the fusion; importantly non-prostate cancer cells 10 (HeLa) are not capable of co-activating this reporter construct with the chimeric fusions.

Figure 4 is a schematic representation of the mechanism of EA1 inhibition.

15 Figure 5 is a bar graph representation of the regulatory effect of EA1/AR Chimera protein on Prostate Specific Promoter.

Figure 6 represents the DNA sequence encoding the EA1/AR (full length) chimeric protein, SEQ ID NO: 1.

20 Figure 7 represents the DNA sequence encoding the EA1/Transactivation Domain (TAD) chimera protein, SEQ ID NO: 2.

25 Figure 8 represents the DNA sequence encoding the EA1/DNA Binding Domain (DBD) chimera protein, SEQ ID NO: 3.

Figures 9 represents the DNA sequence encoding the 12S/AR (full length) chimera protein, SEQ ID NO: 4.

30 Figure 10 represents the DNA sequence encoding the 12S/TAD chimera protein, SEQ ID NO: 5.

Figure 11 represents the DNA sequence encoding the 12S/DBD chimera protein, SEQ ID NO: 6.

Figure 12 represents the amino acid sequence for the 12S/AR (full length) chimera protein, SEQ ID NO: 7.

5 Figure 13 represents the amino acid sequence for the 12S/TAD chimera protein, SEQ ID NO: 8.

Figure 14 represents the amino acid sequence for the 12S/DBD chimera protein, SEQ ID NO: 9.

10 Figure 15 is a schematic representation of some EA1/ AR full length or fragment chimeras of the invention.

Figure 16 represents the cDNA sequence of EA1, SEQ ID NO: 10.

15 Figure 17 represents the cDNA sequence of EA1/TAD chimera, SEQ ID NO: 11.

Figure 18 represents the cDNA sequence of EA1/AR (full length) chimera, SEQ ID NO: 12.

20 Figure 19 represents the cDNA sequence of EA1/AR (C685Y) chimera, SEQ ID NO: 13.

Figure 20 is bar graph representation of the effect of Geldanamycin on Androgen Receptor function in EA1/Ar chimera transfected COS-1 cells.

25 Figure 21 is bar graph representation of the effect of Geldanamycin on Androgen Receptor function in EA1/Ar chimera transfected PC-3 cells.

Figure 22 is a bar graph representation of the effect of androgen agonists and antagonists on the induction of EA1/AR and EA1 (C685Y) in PC3 cells.

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#### Detailed Description of the Invention

We have discovered compositions and methods for enhancing the oncolytic activity of replication-competent, prostate cell-specific adenovirus vectors by modification of the E1A gene product. The target cell-specific replication-competent adenovirus vectors comprise a

chimera of an adenovirus gene essential for replication, specifically the E1A gene, and the Androgen receptor (or a portion thereof) under the transcriptional control of a prostate cell-specific transcriptional regulatory element (TRE). By providing for cell type-specific transcription through the use of one or more cell type-specific TREs, the adenovirus vectors 5 effect prostate-specific cytotoxicity due to selective replication.

The E1A gene inactivates androgen receptor function in prostatic epithelium. Since the androgen receptor is the major regulator of Prostate Specific Antigen promoter activity, conditional replicating adenoviruses, which contain androgen response elements (AREs), are 10 not inducible by androgen. The present invention relates to the generation of E1A chimeric fusion proteins with fragments of the androgen receptor to yield multiple isoforms which are capable of activation of the AREs. The E1A/AR chimeras have several forms. Each form combines different isoforms of the E1A gene product and different domains of the androgen receptor, resulting in a family of chimeras which can be tailored to provide optimal activity of 15 our prostate specific conditional replicating adenoviruses, including the recalcitrant androgen independent prostate cancers often seen in clinical practice.

Compositions of the present invention include, but are not limited to, the chimeric fusion genes in which E1a and the androgen receptor are combined so that the chimera would 20 be capable of recognizing and activating any ARE containing promoter element. Such promoters include the PSA promoter and enhancer, the probasin promoter and the human glandular kallikrein promoter and enhancer. Depending on the chimeric protein used, these constructs can be made active in either androgen dependent or androgen independent disease.

25 In some embodiments, the methods provide for suppressing prostate tumor growth. In other embodiments, the methods are for reducing size and/or extent of a tumor. In other embodiments, the methods are for delaying development of a tumor. In other embodiments, the methods are for treating a neoplasia. In still other embodiments, the methods are for killing tumor cells.

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With respect to all methods described herein, target prostate cells (i.e., neoplastic, proliferative cells) are contacted with an appropriate adenovirus vector described herein (preferably in the form of an adenovirus particle) such that the vector enters the cell and viral replication initiates.

Individuals suitable for treatment by these methods include individuals who have or are suspected of having prostate cancer, including individuals in the early or late stages of the disease, as well as individuals who have previously been treated or are about to undergo treatment (e.g., are in the adjuvant or neoadjuvant setting). Other individuals suitable for the 5 methods described herein are those who are considered high risk for developing a prostate tumor, such as those who have a genetic predisposition to development of a neoplasia and/or who have been exposed to an agent(s) which is correlated with development of a neoplasia.

The presence of prostate cancer and the suitability of the individual for receiving the 10 methods described herein may be determined by any of the techniques known in the art, including diagnostic methods such as imaging techniques, analysis of serum tumor markers, and biopsy.

The various compositions and methods of the invention will be described below. 15 Certain embodiments of the methods use replication-competent prostate cell-specific adenoviral.

Although methods of tumor suppression are exemplified in the discussion below, it is understood that the alternative methods described above are equally applicable and suitable, 20 and that the endpoints of these methods (e.g., efficacy of treatment) are measured using methods standard in the art, including the diagnostic and assessment methods described above.

#### General Techniques:

25 The practice of the present invention will employ, unless otherwise indicated, conventional techniques of molecular biology (including recombinant techniques), microbiology, cell biology, biochemistry and immunology, which are within the scope of those of skill in the art. Such techniques are explained fully in the literature, such as, "Molecular Cloning: A Laboratory Manual", second edition (Sambrook et al., 1989); 30 "Oligonucleotide Synthesis" (M. J. Gait, ed., 1984); "Animal Cell Culture" (R. I. Freshney, ed., 1987); "Methods in Enzymology" (Academic Press, Inc.); "Handbook of Experimental Immunology" (D. M. Weir & C. C. Blackwell, eds.); "Gene Transfer Vectors for Mammalian Cells" (J. M. Miller & M. P. Calos, eds., 1987); "Current Protocols in Molecular Biology" (F.

M. Ausubel et al., eds., 1987); "PCR: The Polymerase Chain Reaction", (Mullis et al., eds., 1994); and "Current Protocols in Immunology" (J. E. Coligan et al., eds., 1991).

For techniques related to adenovirus, see, inter alia, Felgner and Ringold (1989) 5 Nature 337:387-388; Berkner and Sharp (1983) Nucl. Acids Res. 11:6003-6020; Graham (1984) EMBO J. 3:2917-2922; Bett et al. (1993) J. Virology 67:5911-5921; Bett et al. (1994) Proc. Natl. Acad. Sci. USA 91:8802-8806.

**Definitions:**

10 As used herein, the terms "neoplastic cells", "neoplasia", "tumor", "tumor cells", "cancer" and "cancer cells", (used interchangeably) refer to cells which exhibit relatively autonomous growth, so that they exhibit an aberrant growth phenotype characterized by a significant loss of control of cell proliferation. Neoplastic cells can be malignant or benign.

15 As used herein, "suppressing prostate tumor growth" refers to reducing the rate of growth of a tumor, halting tumor growth completely, causing a regression in the size of an existing tumor, eradicating an existing tumor and/or preventing the occurrence of additional tumors upon treatment with the compositions, kits or methods of the present invention. "Suppressing" tumor growth indicates a growth state that is curtailed when compared to 20 growth without contact with, i.e., transfection by, an adenoviral vector. Tumor cell growth can be assessed by any means known in the art, including, but not limited to, directly measuring tumor size, radiographic imaging, utilizing serum biomarkers of disease burden (e.g., serum PSA), determining whether tumor cells are proliferating using a <sup>3</sup>H-thymidine incorporation assay, or counting tumor cells.

25 "Suppressing" prostate tumor cell growth means any or all of the following states: slowing, delaying, and stopping tumor growth, as well as tumor shrinkage.

"Delaying development" of a prostate tumor means to defer, hinder, slow, retard, 30 stabilize, and/or postpone development of the disease. This delay can be of varying lengths of time, depending on the history of the disease and/or individual being treated.

As used herein, "synergy" or "synergistic effect" when referring to combination administration of adenovirus vector and antineoplastic agent and/or radiation means that the

effect of the combination is more than additive when compared to administration of adenovirus vector, antineoplastic agent or radiation alone.

An "adenovirus vector" or "adenoviral vector" (used interchangeably) comprises a 5 polynucleotide construct of the invention. A polynucleotide construct of this invention may be in any of several forms, including, but not limited to, DNA, DNA encapsulated in an adenovirus coat, DNA packaged in another viral or viral-like form (such as herpes simplex, and AAV), DNA encapsulated in liposomes, DNA complexed with polylysine, complexed with synthetic polycationic molecules, conjugated with transferrin, and complexed with 10 compounds such as PEG to immunologically "mask" the molecule and/or increase half-life, and conjugated to a nonviral protein. Preferably, the polynucleotide is DNA. As used herein, "DNA" includes not only bases A, T, C, and G, but also includes any of their analogs or modified forms of these bases, such as methylated nucleotides, internucleotide modifications such as uncharged linkages and thioates, use of sugar analogs, and modified and/or 15 alternative backbone structures, such as polyamides. For purposes of this invention, adenovirus vectors are replication-competent in a target cell.

As used herein, a "transcription response element" or "transcriptional regulatory element", or "TRE" is a polynucleotide sequence, preferably a DNA sequence, which 20 increases transcription of an operably linked polynucleotide sequence in a host cell that allows that TRE to function. A TRE can comprise an enhancer and/or a promoter. A "transcriptional regulatory sequence" is a TRE. A "target cell-specific transcriptional response element" or "target cell-specific TRE" is a polynucleotide sequence, preferably a DNA sequence, which is preferentially functional in a specific type of cell, that is, a target 25 cell. Accordingly, a target cell-specific TRE transcribes an operably linked polynucleotide sequence in a target cell that allows the target cell-specific TRE to function. The term "target cell-specific", as used herein, is intended to include cell type specificity, tissue specificity, developmental stage specificity, and tumor specificity, as well as specificity for a cancerous state of a given target cell. "Target cell-specific TRE" includes cell type-specific and cell 30 status-specific TRE, as well as "composite" TREs. The term "composite TRE" includes a TRE which comprises both a cell type-specific and a cell status-specific TRE. A target cell-specific TRE can also include a heterologous component, including, for example, an SV40 or a cytomegalovirus (CMV) promoter(s). An example of a target cell specific TRE which is tissue specific is a CMV TRE which contains both promoter(s) and enhancer(s).

As described in more detail herein, a target cell-specific TRE can comprise any number of configurations, including, but not limited to, a target cell-specific promoter; and a target cell-specific enhancer; a heterologous promoter and a target cell-specific enhancer; a target cell-specific promoter and a heterologous enhancer; a heterologous promoter and a target cell-specific enhancer; and multimers of the foregoing. The promoter and enhancer components of a target cell-specific TRE may be in any orientation and/or distance from the coding sequence of interest, as long as the desired target cell-specific transcriptional activity is obtained. Transcriptional activation can be measured in a number of ways known in the art (and described in more detail below), but is generally measured by detection and/or quantitation of mRNA or the protein product of the coding sequence under control of (i.e., operably linked to) the target cell-specific TRE. As discussed herein, a target cell-specific TRE can be of varying lengths, and of varying sequence composition. As used herein, the term "cell status-specific TRE" is preferentially functional, i.e., confers transcriptional activation on an operably linked polynucleotide in a cell which allows a cell status-specific TRE to function, i.e., a cell which exhibits a particular physiological condition, including, but not limited to, an aberrant physiological state. "Cell status" thus refers to a given, or particular, physiological state (or condition) of a cell, which is reversible and/or progressive. The physiological state may be generated internally or externally; for example, it may be a metabolic state (such as in response to conditions of low oxygen), or it may be generated due to heat or ionizing radiation. "Cell status" is distinct from a "cell type", which relates to a differentiation state of a cell, which under normal conditions is irreversible. Generally (but not necessarily), as discussed herein, a cell status is embodied in an aberrant physiological state, for example, an oncological state.

Various combinations of transcriptional regulatory elements can be included in a vector. One or more may be heterologous. Further, one or more may have the tissue-specificity. For example, a single transcriptional regulatory element could be used to drive replication by more than one gene essential for replication. This is the case, for example, when the gene product of one of the genes drives transcription of the further gene(s). An example is a heterologous promoter linked to a cassette containing an E1a coding sequence (E1 a promoter deleted) and the entire E1b gene. In such a cascade, only one heterologous transcriptional regulatory element may be necessary. When genes are individually (separately) controlled, however, more than one transcriptional regulatory sequence can be used if more than one such gene is desired to control replication.

A "functional portion" of a target cell-specific TRE is one which confers target cell-specific transcription on an operably linked gene or coding region, such that the operably linked gene or coding region is preferentially expressed in the target cells.

5        By "transcriptional activation" or an "increase in transcription," it is intended that transcription is increased above basal levels in the target cell (i.e., target cell) by at least about 2 fold, preferably at least about 5 fold, preferably at least about 10 fold, more preferably at least about 20 fold, more preferably at least about 50 fold, more preferably at least about 100 fold, more preferably at least about 200 fold, even more preferably at least about 400 fold to about 500 fold, even more preferably at least about 1000 fold. Basal levels are generally the level of activity (if any) in a non-target cell (i.e., a different cell type), or the level of activity (if any) of a reporter construct lacking a target cell-specific TRE as tested in a target cell line.

15       A "functionally-preserved variant" of a target cell-specific TRE is a target cell-specific TRE which differs from another target cell-specific TRE, but still retains target cell-specific transcription activity, although the degree of activation may be altered. The difference in a target cell-specific TRE can be due to differences in linear sequence, arising from, for example, single base mutation(s), addition(s), deletion(s), and/or modification(s) of 20 the bases. The difference can also arise from changes in the sugar(s), and/or linkage(s) between the bases of a target cell-specific TRE. For example, certain point mutations within sequences of TREs have been shown to decrease transcription factor binding and stimulation of transcription. See Blackwood, et al. (1998) Science 281:60-63 and Smith et al. (1997) J. Biol. Chem. 272:27493-27496. One of skill in the art would recognize that some alterations 25 of bases in and around transcription factor binding sites are more likely to negatively affect stimulation of transcription and cell-specificity, while alterations in bases which are not involved in transcription factor binding are not as likely to have such effects. Certain mutations are also capable of increasing TRE activity. Testing of the effects of altering bases may be performed in vitro or in vivo by any method known in the art, such as mobility shift assays, or transfecting vectors containing these alterations in TRE functional and TRE non-functional cells. Additionally, one of skill in the art would recognize that point mutations and 30 deletions can be made to a TRE sequence without altering the ability of the sequence to regulate transcription.

As used herein, a TRE derived from a specific gene is referred to by the gene from which it was derived and is a polynucleotide sequence which regulates transcription of an operably linked polynucleotide sequence in a host cell that expresses said gene. For example, as used herein, a "human glandular kallikrein transcriptional regulatory element", or "hK2-TRE" is a polynucleotide sequence, preferably a DNA sequence, which increases transcription of an operably linked polynucleotide sequence in a host cell that allows an hK2-TRE to function, such as a cell (preferably a mammalian cell, even more preferably a human cell) that expresses androgen receptor, such as a prostate cell. An hK2-TRE is thus responsive to the binding of androgen receptor and comprises at least a portion of an hK2 promoter and/or an hK2 enhancer (i.e., the ARE or androgen receptor binding site).

As used herein, a "probasin (PB) transcriptional regulatory element", or "PB-TRE" is a polynucleotide sequence, preferably a DNA sequence, which selectively increases transcription of an operably-linked polynucleotide sequence in a host cell that allows a PB-TRE to function, such as a cell (preferably a mammalian cell, more preferably a human cell, even more preferably a prostate cell) that expresses androgen receptor. A PB-TRE is thus responsive to the binding of androgen receptor and comprises at least a portion of a PB promoter and/or a PB enhancer (i.e., the ARE or androgen receptor binding site).

As used herein, a "prostate-specific antigen (PSA) transcriptional regulatory element", or "PSA-TRE", or "PSE-TRE" is a polynucleotide sequence, preferably a DNA sequence, which selectively increases transcription of an operably linked polynucleotide sequence in a host cell that allows a PSA-TRE to function, such as a cell (preferably a mammalian cell, more preferably a human cell, even more preferably a prostate cell) that expresses androgen receptor. A PSA-TRE is thus responsive to the binding of androgen receptor and comprises at least a portion of a PSA promoter and/or a PSA enhancer (i.e., the ARE or androgen receptor binding site).

As used herein, a target cell-specific TRE can comprise any number of configurations, including, but not limited to, a target cell-specific promoter; a target cell-specific enhancer; a target cell-specific promoter and a target cell-specific enhancer; a target cell-specific promoter and a heterologous enhancer; a heterologous promoter and a target cell-specific enhancer; and multimers of the foregoing. The promoter and enhancer components of a target cell-specific TRE may be in any orientation and/or distance from the coding sequence of

interest, as long as the desired target cell-specific transcriptional activity is obtained. Transcriptional activation can be measured in a number of ways known in the art (and described in more detail below), but is generally measured by detection and/or quantitation of mRNA or the protein product of the coding sequence under control of (i.e., operably linked to) the target cell-specific TRE.

A "multicistronic transcript" refers to an mRNA molecule which contains more than one protein coding region, or cistron. A mRNA comprising two coding regions is denoted a "bicistronic transcript." The "5'-proximal" coding region or cistron is the coding region whose translation initiation codon (usually AUG) is closest to the 5'-end of a multicistronic mRNA molecule. A "5'-distal" coding region or cistron is one whose translation initiation codon (usually AUG) is not the closest initiation codon to the 5' end of the mRNA. The terms "5'-distal" and "downstream" are used synonymously to refer to coding regions that are not adjacent to the 5' end of a mRNA molecule.

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As used herein, "co-transcribed" means that two (or more) coding regions of polynucleotides are under transcriptional control of single transcriptional control element.

A "gene" refers to a coding region of a polynucleotide. A "gene" may or may not include non-coding sequences and/or regulatory elements.

"Replicating preferentially", as used herein, means that the adenovirus replicates more in a target cell than a non-target cell. Preferably, the adenovirus replicates at a significantly higher rate in target cells than non target cells; preferably, at least about 2-fold higher, preferably, at least about 5-fold higher, more preferably, at least about 10-fold higher, still more preferably at least about 50-fold higher, even more preferably at least about 100-fold higher, still more preferably at least about 400- to 500-fold higher, still more preferably at least about 1000-fold higher, most preferably at least about  $1 \times 10^6$  higher. Most preferably, the adenovirus replicates solely in the target cells (that is, does not replicate or replicates at a very low levels in non-target cells).

As used herein, the term "vector" refers to a polynucleotide construct designed for transduction/transfection of one or more cell types. Vectors may be, for example, "cloning vectors" which are designed for isolation, propagation and replication of inserted nucleotides,

"expression vectors" which are designed for expression of a nucleotide sequence in a host cell, or a "viral vector" which is designed to result in the production of a recombinant virus or virus-like particle, or "shuttle vectors", which comprise the attributes of more than one type of vector.

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The terms "polynucleotide" and "nucleic acid", used interchangeably herein, refer to a polymeric form of nucleotides of any length, either ribonucleotides and/or deoxyribonucleotides. These terms include a single-, double- or triple-stranded DNA, genomic DNA, cDNA, RNA, DNA-RNA hybrid, or a polymer comprising purine and pyrimidine bases, or other natural, chemically, biochemically modified, non-natural or derivatized nucleotide bases. The backbone of the polynucleotide can comprise sugars and phosphate groups (as may typically be found in RNA or DNA), or modified or substituted sugar or phosphate groups. Alternatively, the backbone of the polynucleotide can comprise a polymer of synthetic subunits such as phosphoramidates and thus can be a oligodeoxynucleoside phosphoramidate (P-NH<sub>2</sub>) or a mixed phosphoramidate-phosphodiester oligomer. Peyrottes et al. (1996) Nucleic Acids Res. 24: 1841-8; Chaturvedi et al. (1996) Nucleic Acids Res. 24: 2318-23; Schultz et al. (1996) Nucleic Acids Res. 24: 2966-73. A phosphorothioate linkage can be used in place of a phosphodiester linkage. Braun et al. (1988) J. Immunol. 141: 2084-9; Latimer et al. (1995) Molec. Immunol. 32: 1057-1064. In addition, a double-stranded polynucleotide can be obtained from the single stranded polynucleotide product of chemical synthesis either by synthesizing the complementary strand and annealing the strands under appropriate conditions, or by synthesizing the complementary strand *de novo* using a DNA polymerase with an appropriate primer. Reference to a polynucleotide sequence (such as referring to a SEQ ID NO) also includes the complement sequence.

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The following are non-limiting examples of polynucleotides: a gene or gene fragment, exons, introns, mRNA; tRNA; rRNA, ribozymes, cDNA, recombinant polynucleotides, branched polynucleotides, plasmids, vectors, isolated DNA of any sequence, isolated RNA of any sequence, nucleic acid probes, and primers. A polynucleotide may comprise modified nucleotides, such as methylated nucleotides and nucleotide analogs, uracyl, other sugars and linking groups such as fluororibose and thioate, and nucleotide branches. The sequence of nucleotides may be interrupted by non-nucleotide components. A polynucleotide may be further modified after polymerization, such as by conjugation with a labeling component.

Other types of modifications included in this definition are caps, substitution of one or more of the naturally occurring nucleotides with an analog, and introduction of means for attaching the polynucleotide to proteins, metal ions, labeling components, other polynucleotides, or a solid support. Preferably, the polynucleotide is DNA. As used herein, "DNA" includes not only bases A, T, C, and G, but also includes any of their analogs or modified forms of these bases, such as methylated nucleotides, internucleotide modifications such as uncharged linkages and thioates, use of sugar analogs, and modified and/or alternative backbone structures, such as polyamides.

10       A polynucleotide or polynucleotide region has a certain percentage (for example, 80%, 85%, 90%, or 95%) of "sequence identity" to another sequence means that, when aligned, that percentage of bases are the same in comparing the two sequences. This alignment and the percent homology or sequence identity can be determined using software programs known in the art, for example those described in Current Protocols in Molecular 15      Biology (F. M. Ausubel et al., eds., 1987) Supplement 30, section 7.7.18. A preferred alignment program is ALIGN Plus (Scientific and Educational Software, Pennsylvania), preferably using default parameters, which are as follows: mismatch=2; open gap=0; extend gap=2.

20       "Under transcriptional control" is a term well understood in the art and indicates that transcription of a polynucleotide sequence, usually a DNA sequence, depends on its being operably (operatively) linked to an element which contributes to the initiation of, or promotes, transcription. "Operably linked" refers to a juxtaposition wherein the elements are in an arrangement allowing them to function.

25       The term "enhancer" is used according to its art-recognized meaning. It is intended to mean a sequence found in eukaryotes and certain eukaryotic viruses which can increase transcription from a gene when located (in either orientation) up to several kilobases from the gene being studied. These sequences usually act as enhancers when on the 5' side (upstream) 30      of the gene in question. However, some enhancers are active when placed on the 3' side (downstream) of the gene. In some cases, enhancer elements can activate transcription from a gene with no (known) promoter.

The term "functional inactivation" is intended to mean a genetic lesion that prevents the normal activity of a gene product. Thus, functional inactivation could result from a mutation in the gene encoding the gene product. Such a lesion includes insertions, deletions, and base changes. Alternatively, functional inactivation may occur by the abnormal interaction of the normal gene product with one or more other cellular gene products which bind to or otherwise prevent the functional activity of said gene product. Thus, the gene product may be a protein produced from a normal gene but which cannot perform its ordinary and normal function because of an interaction with a second factor.

10 The term "gene essential for replication" refers to a genetic sequence whose transcription is required for the vector to replicate in the target cell.

15 The term "gene product" is intended to mean DNA, RNA, protein, peptides, or viral particles. Thus, the distribution, for the purposes of the invention, is of any of these components.

20 The term "heterologous" means a DNA sequence not found in the native vector genome. With respect to a "heterologous transcriptional regulatory sequence", "heterologous" indicates that the transcriptional regulatory sequence is not naturally ligated to the DNA sequence for the gene essential for replication of the vector.

25 The term "promoter" is used according to its art-recognized meaning. It is intended to mean the DNA region, usually upstream to the coding sequence of a gene or operon, which binds RNA polymerase and directs the enzyme to the correct transcriptional start site.

30 The term "replication" means duplication of a vector. This duplication, in the case of viruses, can occur at the level of nucleic acid, or at the level of infectious viral particle. In the case of DNA viruses, replication at the nucleic acid level comprises DNA replication. In the case of RNA viruses, nucleic acid replication comprises replication into plus or minus strand (or both). In the case of retroviruses, replication at the nucleic acid level includes the production of cDNA as well as the further production of RNA viral genomes. The essential feature is the generation of nucleic acid copies of the original viral vector. However, replication also includes the formation of infectious DNA or RNA viral particles. Such

particles may successively infect cells in a given target tissue, thus distributing the vector through all or a significant portion of the target tissue.

The term "replication-conditional vector" refers to a vector which, when introduced 5 into a tissue, will not replicate unless a transcriptional regulatory sequence in that vector is activated or derepressed in that tissue. That is, replication depends upon transcription by means of that transcriptional regulatory sequence. Such a vector is replication-conditional as described because it has been modified in the following manner. A gene that is essential for replication has been modified by replacing the transcriptional regulatory sequence on which 10 transcription of that gene normally depends with a heterologous transcriptional regulatory sequence. This transcriptional regulatory sequence depends upon the presence of transcriptional regulatory factors or the absence of transcriptional regulatory inhibitors. The presence of these factors in a given tissue or the absence of such inhibitors in a given tissue provides the replication-conditionality. Accordingly, the native transcriptional regulatory 15 sequence may be replaced with the heterologous transcriptional regulatory sequence. Alternatively, the native transcriptional regulatory sequence may be disabled or rendered dysfunctional by partial removal (deletion) or other mutation (one or more base changes, insertions, inversions, etc.).

20 The gene sequence may be a coding sequence. It may contain one or more open reading frames, as well as intron sequences. However, such a sequence is not limited to a coding sequence, but includes sequences that are transcribed into RNA, which RNA is itself essential for vector replication. The essential feature is that the transcription of the gene sequences does not depend on the native transcriptional regulatory sequences.

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The term "silencer," used in its art-recognized sense, means a sequence found in eucaryotic viruses and eucaryotes which can decrease or silence transcription of a gene when located within several kilobases of that gene.

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The term "tissue-specific" is intended to mean that the transcriptional regulatory sequence to which the gene essential for replication is operably linked functions specifically in that tissue so that replication proceeds in that tissue. This can occur by the presence in that tissue, and not in non-target tissues, of positive transcription factors that activate the transcriptional regulatory sequence. It can also occur by the absence of transcription

inhibiting factors that normally occur in non-target tissues and prevent transcription as a result of the transcription regulatory sequence. Thus, when transcription occurs, it proceeds into the gene essential for replication such that in that target tissue, replication of the vector and its attendant functions occur.

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As described herein, tissue specificity is particularly relevant in the treatment of the abnormal counterpart of a normal tissue. Such counterparts include, but are not limited to, cancerous prostate tissue and normal prostate tissue. Tissue specificity also includes the presence of an abnormal tissue type interspersed with normal tissue of a different tissue type, 10 as for example in the case of metastases of prostate cancer, and the like, into tissue such as liver. In this case, the target tissue is the abnormal tissue, and tissue specificity reflects the restriction of vector replication to the abnormal tissue interspersed in the normal tissue. It is also to be understood that tissue specificity, in the context of treatment, is particularly relevant *in vivo*. However, as described herein, *ex vivo* treatment and tissue replacement also 15 falls within the concept of tissue specificity according to the present invention.

An "E3 region" (used interchangeably with "E3") is a term well understood in the art and means the region of the adenoviral genome that encodes the E3 products (discussed herein). Generally, the E3 region is located between about 28583 and 30470 of the adenoviral 20 genome. The E3 region has been described in various publications, including, for example, Wold et al. (1995) *Curr. Topics Microbiol Immunol.* 199:237-274.

A "portion" of the E3 region means less than the entire E3 region, and as such includes polynucleotide deletions as well as polynucleotides encoding one or more 25 polypeptide products of the E3 region. As used herein, "cytotoxicity" is a term well understood in the art and refers to a state in which a cell's usual biochemical or biological activities are compromised (i.e., inhibited). These activities include, but are not limited to, metabolism; cellular replication; DNA replication; transcription; translation; and uptake of molecules. "Cytotoxicity" includes cell death and/or cytolysis. Assays are known in the art 30 which indicate cytotoxicity, such as dye exclusion, <sup>3</sup>H-thymidine uptake, and plaque assays.

An "E1B 19-kDa region" (used interchangeably with "E1B 19-kDa genomic region") refers to the genomic region of the adenovirus E1B gene encoding the E1B 19-kDa product. According to wild-type Ad5, the E1B 19-kDa region is a 261 bp region located between

nucleotide 1714 and nucleotide 2244. The E1B 19-kDa region has been described in, for example, Rao et al., Proc. Natl. Acad. Sci. USA, 89:7742-7746. The present invention encompasses deletion of part or all of the E1B 19-kDa region as well as embodiments wherein the E1B 19-kDa region is mutated, as long as the deletion or mutation lessens or 5 eliminates the inhibition of apoptosis associated with E1B-19 kDa.

The term "selective cytotoxicity", as used herein, refers to the cytotoxicity conferred by an adenovirus vector of the present invention on a cell which allows or induces a target cell-specific TRE to function (a target cell) when compared to the cytotoxicity conferred by 10 an adenoviral vector of the present invention on a cell which does not allow a target cell-specific TRE to function (a non-target cell). Such cytotoxicity may be measured, for example, by plaque assays, by reduction or stabilization in size of a tumor comprising target cells, or the reduction or stabilization of serum levels of a marker characteristic of the tumor cells, or a tissue-specific marker, e.g., a cancer marker.

15

In the context of adenovirus, a "heterologous polynucleotide" or "heterologous gene" or "transgene" is any polynucleotide or gene that is not present in wild-type adenovirus. Preferably, the transgene will also not be expressed or present in the target cell prior to introduction by the adenovirus vector.

20

In the context of adenovirus, a "heterologous" promoter or enhancer is one which is not associated with or derived from an adenovirus gene.

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In the context of adenovirus, an "endogenous" promoter, enhancer, or TRE is native to or derived from adenovirus. In the context of promoter, an "inactivation" means that there is a mutation of or deletion in part or all of the endogenous promoter, i.e., a modification or alteration of the endogenous promoter, such as, for example, a point mutation or insertion, which disables the function of the promoter.

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In the context of a target cell-specific TRE, a "heterologous" promoter or enhancer is one which is derived from a gene other than the gene from which a reference target cell-specific TRE is derived.

A "host cell" includes an individual cell or cell culture which can be or has been a recipient of an adenoviral vector(s) of this invention. Host cells include progeny of a single host cell, and the progeny may not necessarily be completely identical (in morphology or in total DNA complement) to the original parent cell due to natural, accidental, or deliberate mutation and/or change. A host cell includes cells transfected or infected in vivo or in vitro with an adenoviral vector of this invention.

"Replication" and "propagation" are used interchangeably and refer to the ability of an adenovirus vector of the invention to reproduce or proliferate. These terms are well understood in the art. For purposes of this invention, replication involves production of adenovirus proteins and is generally directed to reproduction of adenovirus. Replication can be measured using assays standard in the art and described herein, such as a burst assay or plaque assay. "Replication" and "propagation" include any activity directly or indirectly involved in the process of virus manufacture, including, but not limited to, viral gene expression; production of viral proteins, nucleic acids or other components; packaging of viral components into complete viruses; and cell lysis.

"Androgen receptor," or AR, as used herein refers to a protein whose function is to specifically bind to androgen and, as a consequence of the specific binding, recognize and bind to an androgen response element (ARE), following which the AR is capable of regulating transcriptional activity. The AR is a nuclear receptor that, when activated, binds to cellular androgen-responsive element(s). In normal cells the AR is activated by androgen, but in non-normal cells (including malignant cells) the AR may be activated by non-androgenic agents, including hormones other than androgens. Encompassed in the term "androgen receptor" are mutant forms of an androgen receptor, such as those characterized by amino acid additions, insertions, truncations and deletions, as long as the function is sufficiently preserved. Mutants include androgen receptors with amino acid additions, insertions, truncations and deletions, as long as the function is sufficiently preserved. In this context, a functional androgen receptor is one that binds both androgen and, upon androgen binding, an ARE.

A polynucleotide sequence that is "depicted in" a SEQ ID NO means that the sequence is present as an identical contiguous sequence in the SEQ ID NO. The term

encompasses portions, or regions of the SEQ ID NO as well as the entire sequence contained within the SEQ ID NO.

A "biological sample" encompasses a variety of sample types obtained from an individual and can be used in a diagnostic or monitoring assay. The definition encompasses blood and other liquid samples of biological origin, solid tissue samples such as a biopsy specimen or tissue cultures or cells derived therefrom, and the progeny thereof. The definition also includes samples that have been manipulated in any way after their procurement, such as by treatment with reagents, solubilization, or enrichment for certain components, such as proteins or polynucleotides. The term "biological sample" encompasses a clinical sample, and also includes cells in culture, cell supernatants, cell lysates, serum, plasma, biological fluid, and tissue samples.

An "individual" is a vertebrate, preferably a mammal, more preferably a human. Mammals include, but are not limited to, farm animals, sport animals, rodents, primates, and pets

An "effective amount" is an amount sufficient to effect beneficial or desired results, including clinical results. An effective amount can be administered in one or more administrations. For purposes of this invention, an effective amount of an adenoviral vector is an amount that is sufficient to palliate, ameliorate, stabilize, reverse, slow or delay the progression of the disease state.

A given TRE is "derived from" a given gene if it is associated with that gene in nature.

"Expression" includes transcription and/or translation.

As used herein, the term "comprising" and its cognates are used in their inclusive sense; that is, equivalent to the term "including" and its corresponding cognates.

"A," "an" and "the" include plural references unless the context clearly dictates otherwise.

**Adenoviral E1A Mediated Viral Replication:**

Type 5 adenoviruses consist of roughly 36 kilobases encoding over 38 gene products. For convenience, the genes are categorized into early (E1-E4) and late (L1-L5) regions. The initiation of the replicative life cycle requires the E1 region, which encodes the E1A and E1B genes. E1A interacts with many proteins including the CREB binding protein (CBP), p300, pRB and related proteins p130 and p107, p400, TRAP, p60/cyclin A, p33 cdk2, BS69, CtBP, the TATA binding protein (TBP) and various other components of the TFIID complex (Virus Res 1995; 35:113-21, Biochem Cell Biol 1997; 75: 95-102). These interactions account for many of the elucidated functions of E1A, including, but not limited to: chromatin remodeling, regulation of histone acetyltransferase, transcriptional activation, transcriptional repression, induction of DNA synthesis, and suppression of differentiation (Nat Rev Mol Cell Biol 2002; 3: 441-52). In the appropriately susceptible host cell, E1A and E1B coordinate viral replication, with passive lysis of the cell and release of the viral progeny. Since the adenoviral DNA is maintained completely in the episomal state, infected cells either undergo apoptosis in an abortive infection, or viral replication followed by lysis.

Immediately after cellular infection, the E1 genes are transcribed and bind to a host of protein complexes. Although the order of binding is unclear, several key features are known. Firstly, E1A binds to p400 and TRAP, which have homology to the Swi/Snf family of ATP-dependent chromatin remodeling factors. These factors aid in regulating gene expression by allowing access to "euchromatin". E1A also binds to at least three protein complexes known to have significant histone acetyl transferase activity: p300, CREB binding protein (CBP) and PCAF. Active acetylation of certain histones (e.g., H3 and H4) is known to facilitate DNA unwinding and enhance gene expression by allowing easier access to transcription factors and co-activators. Hence, host transcription and replication is strongly down regulated and adenoviral transcription and replication is strongly up regulated.

E1A is known to both activate and inhibit certain trans-gene expression. There appear to be at least two different mechanisms by which this can occur (Nat Rev Mol Cell Biol 2002; 3: 441-52; Oncogene 2001; 20: 7824-35): (1) sequestration of chromatin remodeling/HAT complexes required by certain transcription factors; (2) active inhibition of certain trans-genes by direct binding of E1A with the transcription factor and the inhibitory complex C-terminal binding protein (CtBP). CBP/p300 and PCAF are not required for expression of all genes, but rather only a small subset of genes regulated by transcription

factors thought to be most important in differentiation. Two-prostate specific oncolytic adenoviruses, CN706 (also known as CG7060) and CV787 (also known as CG7870) are unable to undergo androgen induction for enhanced oncolytic activity (Cancer Res 1999; 59:4200-3). This finding is surprising given that they are regulated by ARE containing 5 promoters. In contrast, generation of an estrogen receptor dependent conditionally replication competent adenovirus, by placing a portion of the pS2 promoter upstream of the E1A gene, resulted in a viral construct whose oncolytic activity was markedly enhanced by estrogen and antagonized by anti-estrogen (Hum Gene Ther 2000; 2009-24). In that analysis, the addition of estrogen caused a marked increase in the E1A expression and this increase 10 precisely correlated with the viral oncolytic activity in ER positive cells. Evolving data regarding E1A protein interactions suggest that E1A is able to interact directly with some members of the nuclear receptor family. E1A interaction with RAR- $\beta$  augments its ability to stimulate ligand dependent activation of the RAR- $\beta$ 2 promoter (Mol Cell Biol 1995; 10: 19-25); while in contrast, E1A interaction with RXR retinoic acid receptors obliterates retinoic 15 acid(RA) induction of differentiation genes (Oncogene 1995; 10: 19-25). Notably, E1A directly binds to RAR- $\beta$  (in the CR3 region), but does not bind RXR. Simply stated, E1A is capable of either up-regulating or down-regulating retinoic acid responsive target genes dependent on the particular target gene promoter and the nature of the retinoic acid receptor complex (RXR versus RAR). Given these findings, we hypothesized that the reason we did 20 not see androgen induction in the early prostate specific oncolytic vectors was because E1A was specifically inhibiting AR activity, a finding we later confirmed.

#### Androgen Receptor Structure and Function:

The androgen receptor is a member of the nuclear receptor super-family, which is a 25 group of ligand-inducible transcription factors. Like other steroid receptors, the structure of the androgen receptor is divided into three distinct, modular domains: the amino terminal domain, the DNA binding domain and the carboxy-terminal ligand binding domain (Vitam Horm 1994; 49: 383-432). Binding of the steroid ligand results in activation of the androgen receptor and dissociation from the chaperonin complex (Hsp90, Hsp70, Hip, p60, p23, 30 FKBP51, FKBP52 and Cyp40). Subsequently, the receptor undergoes dimerization, nuclear translocation and subsequent target gene activation (Annu Rev Med 1995; 46: 443-53). Binding of androgen to the ligand-binding domain is required for activation; however, deletion of the entire ligand-binding domain can lead to a constitutively active androgen

receptor. Interestingly, point mutations in the ligand-binding domain can result in altered steroid specificity. The best characterized of these is a single point mutation resulting in a threonine to alanine substitution at codon 868 identified in the LNCaP cell line (J Steroid Biochem Mol Biol 1994; 49: 341-6), resulting in an AR which can be induced by  
5 progesterone as well as androgen. Conversion of codon 685 from a cysteine to a tyrosine results in an androgen receptor mutant which can be strongly activated with the nonsteroidal anti-androgen bicalutamide (Endocrine 2000; 12: 69-76). Once the androgen receptor has achieved translocation into the nucleus, it must coordinate binding to a number of associated co-activators and co-repressors, which subsequently regulate gene expression. As the number  
10 of potential co-regulators clearly exceeds the capacity for direct interaction by a single receptor, the most likely scenario is that transcriptional activation by AR involves multiple factors that act in both a sequential and concomitant manner to reorganize chromatin templates, in a manner analogous to the adenoviral E1A. The precise timing and sequence of binding of these factors remains to be elucidated, however, one can generally break down the  
15 processes empirically into chromatin/nucleosomal remodeling (an energy dependent process), histone acetyltransferase activity, and subsequent recruitment of TATA binding protein associated factors (TAFs) all of which promote an increased rate of gene transcription by RNA polymerase II.

20 N-terminal deletions in region of codons 46-408 results in dominant negative suppression of hormone inducible transgene activation, indicating that the major co-activator functions require an interaction within that site. The major transactivating segment is referred to as the activator function-1 (AF-1), which is required for interaction with SRC-1. Maximal co-activation however, requires an interaction with the ligand binding domain LXXLL motif  
25 at the AF-2 site. The androgen receptor interacts with chromatin remodeling complexes in an ATP dependent fashion and this may be among the earliest steps in the ultimate regulation of certain target genes. Such complexes include the multisubunit human SWI-SNF complex, which has been shown to remodel mono-and polynucleosome templates. Since condensed chromatin renders genes inaccessible for transcription, the combination of a steroid receptor  
30 along with the SWI-SNF complex formation appears to be critical for appropriate nucleosomal remodeling to allow appropriate target genes to be accessible for gene regulation. Once the receptor SWI-SNF complex has successfully "opened" the structure of the chromatin to allow transcriptional regulation, the androgen receptor must interact with the HAT complexes p300/CBP and PCAF as well as a series of co-activators. Among the most

important co-activators is the SRC-1, which has mild HAT activity and seems to be required for optimal stimulation of steroid dependent transcription. Additional factors include SRA (a structural RNA necessary for the coactivator complex to function optimally), and p160, which appears to be required for hormone dependent activation and appears to directly

5 interact with the polyglutamine repeats found in the amino terminal transactivation domain. Maximal androgen receptor mediated transactivation requires an additional interaction with a large multi-protein complex referred to as TRAP (for thyroid receptor associated complex) /DRIP (for vitamin D Receptor interacting proteins) /ARC for activator-recruited cofactor complex. Though initially described for their interactions with the thyroid hormone receptor

10 and vitamin D receptor, this complex is utilized by a variety of transcription factors, which stimulate RNA Pol II transcription, including the androgen receptor. In summary, binding of steroid ligand by the androgen receptor results in dissociation from the chaperonin complex, dimerization, post-translational modification, nuclear translocation, interactions with chromatin remodeling complexes, interactions with HAT complexes, interactions with

15 numerous coactivator complexes, and interactions with complexes directly involved in initiation of transcription from the TFIID complex. As can be seen, there are distinct similarities between E1A interactions and the androgen receptor interactions, which led us to question whether E1A might differentially regulate AR action.

20 **EA1/ Androgen Receptor Chimeras:**

The invention also provides chimeric or fusion nucleotides of the E1A and Androgen Receptor genes (or portions thereof) described herein. As used herein, a “chimeric gene” or “fusion gene” comprises genes of the invention operatively linked to heterologous genes sequences. “Heterologous gene sequences” includes a gene or polynucleotide having a sequence corresponding to a gene which is not substantially homologous to gene of the invention. Within the fusion gene, the term “operatively linked” is intended to indicate that the gene of the invention and the heterologous gene sequences are fused in-frame to each other. The heterologous gene sequences can be fused to the N-terminus or C-terminus of the corresponding peptide of the invention.

25

In various embodiments, the chimera is comprised of sequences of the E1A gene and fragments of the Androgen receptor gene, such as the transactivation domain and the DNA Binding Domain. In another embodiment, there is a single point mutation the full length amino acid sequence of the Androgen receptor in the E1A/AR chimeric protein. The single

point mutation occurs at the codon 685 position, where cysteine is replaced by tyrosine, which results in a chimera activated by both androgens and nonsteroidal anti-androgens.

Preferably, a chimeric or fusion gene of the invention is produced by standard recombinant DNA techniques. For example, DNA fragments coding for the different peptide sequences are ligated together in-frame in accordance with conventional techniques, for example by employing blunt-ended or stagger-ended termini for ligation, restriction enzyme digestion to provide for appropriate termini, filling-in of cohesive ends as appropriate, alkaline phosphatase treatment to avoid undesirable joining, and enzymatic ligation. In another embodiment, the fusion gene can be synthesized by conventional techniques including automated DNA synthesizers. Alternatively, PCR amplification of gene fragments can be carried out using anchor primers which give rise to complementary overhangs between two consecutive gene fragments which can subsequently be annealed and reamplified to generate a chimeric gene sequence (see, for example, Current Protocols in Molecular Biology, eds. Ausubel et al. John Wiley & Sons: 1992). Moreover, many expression vectors are commercially available that already encode a fusion moiety (e.g., a GST polypeptide). A peptide-encoding nucleic acid can be cloned into such an expression vector such that the fusion moiety is linked in-frame to the peptide.

Co-transfection assays of prostate cancer cell lines LNCaP and LAPC4 CMV-E1A-AR expressing plasmid or CMV-E1A expressing plasmid and luciferase reporter plasmid controlled by prostate specific promoter demonstrate that the expression of E1A completely inhibits the androgen receptor induction of the prostate specific promoter. This inhibitory effect of E1A on androgen receptor can be reversed by expression of E1A-AR. A similar effect can be observed in the assay using adenovirus infection, i.e. E1A expressing Ad5 can down-regulate the androgen receptor, and this inhibitory effect could be counteracted by E1A-AR expressing Ad5.

Other data demonstrate that either wild type E1A or its isoform 12S have the same inhibitory effect on the androgen receptor. Either full-length of androgen receptor or its truncated forms TAD-DBD (transactivation domain-DNA binding domain), or DBD (DNA binding domain) is able to counteract the inhibitory effect of E1A on androgen receptor. Each form of androgen receptor can display a unique profile of its specificity and potency in regulating the activity of the prostate specific promoter when it is in an E1A-AR fusion

format. Different combinations of E1A-AR expressing adenoviral vectors could meet the need in the clinic of treatment for different types of prostate cancer (e.g. hormone-dependent cells or hormone-refractory cells). These cancer cell-tailored vectors will achieve a maximum therapeutic effect in the clinic. The nature of our vectors makes them different from and  
5 much better than the existing technologies for developing conditionally replicating adenoviruses in prostate cancer.

The adenoviral vector compositions and methods of the present invention and described herein are generally replication-competent, prostate-cell specific adenoviral vector  
10 comprising a chimera of an adenovirus gene essential for replication and the Androgen Receptor (and portions thereof) under transcriptional control of a TRE, embodiments of which are described infra. In one embodiment, the gene essential for replication in the adenoviral vector is an early gene, specifically E1A. In another embodiment E1A/AR chimera genes are under transcriptional control of non-identical (or heterologous) TREs. In  
15 some embodiments, the adenovirus vector E3 region is deleted.

The choice of adenoviral vector is primarily determined by the identity of the target cells, in this case prostate cells and therefore the type of cancer to be treated. As explained below in detail, an adenoviral vector comprising a PSA-TRE, PB-TRE, or hK2-TRE would  
20 preferentially replicate in prostate cells.

For example, with respect to treatment of prostate tumors, a replication-competent adenovirus in which a gene essential for replication, preferably one or more early genes, is under transcriptional control of a prostate specific TRE.  
25

In some embodiments of the adenovirus vector, E1A is under transcriptional control of a prostate specific TRE. An example of a suitable prostate specific replication-competent adenoviral vector is one comprising probasin (PB)-TRE controlling transcription of E1A, and PSE-TRE controlling transcription of E1B, such as CG7870.

30 In some embodiments, a prostate specific adenoviral vector comprising a chimera of E1A and the Androgen receptor under transcriptional control of two non-identical prostate specific TREs. In some embodiments, the prostate specific TRE controlling transcription of E1A/Androgen Receptor chimera are heterologous (i.e., of different sequence) with respect to

each other. In some embodiments, the prostate specific TRE controlling transcription of the E1A/Androgen Receptor chimera is derived from probasin (PB) and prostate specific antigen (PSA).

5       The adenoviral vectors used in the methods described herein are replication-competent target-cell specific adenoviral vectors comprising an adenovirus gene, preferably a gene essential for replication under transcriptional control of a target cell specific TRE. The vector may or may not include an E3 region. In other embodiments, an adenovirus vector is a replication competent, target cell specific vector comprising E1B, wherein E1B may have a  
10      deletion of part or the entire 19-kDa region.

The adenovirus vectors used in this invention replicate preferentially in TRE functional cells referred to herein as prostate cells. This replication preference is indicated by comparing the level of replication (i.e., titer) in prostate cells in which the TRE is active to  
15      the level of replication in cells in which the TRE is not active (i.e., a non-target cell). The replication preference is even more significant, as the adenovirus vectors used in the invention actually replicate at a significantly lower rate in TRE non-functional cells than wild type virus. Comparison of the adenovirus titer of a target cell to the titer of a TRE inactive cell type provides a key indication that the overall replication preference is enhanced due to  
20      the replication in target cells as well as depressed replication in non-target cells. This is especially useful in the cancer context, in which targeted cell killing is desirable. The TRE's preferably control genes necessary for replication, where the gene(s) necessary for replication is an early gene(s) of the adenovirus, preferentially the E1A gene.

25       Runaway infection is prevented due to the cell-specific requirements for viral replication. Without wishing to be bound by any particular theory, production of adenovirus proteins can serve to activate and/or stimulate the immune system, either generally or specifically toward target cells producing adenoviral proteins which can be an important consideration in the cancer context, where individuals are often moderately to severely  
30      immunocompromised.

In particular embodiments, the adenoviral vector may be a replication-competent target-cell specific adenoviral vector where the vector comprises an adenoviral gene. In one

embodiment, the adenoviral gene is essential for replication and is under transcriptional control of a target cell-specific TRE.

In certain embodiments, the adenoviral vector may be a replication-competent target-cell specific adenoviral vector wherein the gene essential for replication is an early gene. In other embodiments the gene essential for replication may be a late gene.

In some embodiments of the adenovirus vector, E1A has a mutation in or deletion of its endogenous promoter. In some embodiments, E1A has a mutation in or deletion of its endogenous enhancer.

In particular preferred embodiments, the target cell specific adenoviral vector is specific for target cells of the prostate.

In certain preferred embodiments, the adenoviral gene(s) essential for replication is under the control of a TRE(s) such as, but not limited to PB-TRE, PSA-TRE, hK2-TRE, as described herein.

#### Transcriptional Response Elements (TREs):

The adenovirus vectors of the invention comprise target cell specific TREs which direct preferential expression of an operatively linked gene (or genes) in a particular target cell. A TRE can be tissue-specific, tumor-specific, developmental stage-specific, cell status specific, etc., depending on the type of cell present in the prostate tumor.

Cell- and tissue-specific transcriptional regulatory elements, as well as methods for their identification, isolation, characterization, genetic manipulation and use for regulation of operatively linked coding sequences, are well known in the art. A TRE can be derived from the transcriptional regulatory sequences of a single gene, or sequences from different genes can be combined to produce a functional TRE. A cell-specific TRE is preferentially functional in a limited population (or type) of cells, e.g., prostate cells. Accordingly, in some embodiments, the TRE used is preferentially functional in any of the prostate cell types.

As is known in the art, activity of TREs can be inducible. Inducible TREs generally exhibit low activity in the absence of inducer, and are up-regulated in the presence of inducer.

Inducers include, for example, nucleic acids, polypeptides, small molecules, organic compounds and/or environmental conditions such as temperature, pressure or hypoxia. Inducible TREs may be preferred when expression is desired only at certain times or at certain locations, or when it is desirable to titrate the level of expression using an inducing agent. For example, transcriptional activity from the PSA-TRE, PB-TRE and hK2-TRE is inducible by androgen, as described herein and in PCT/US98/04080. Accordingly, in one embodiment of the present invention, an adenovirus vector comprises an inducible heterologous TRE.

TRE multimers are also useful in the disclosed vectors. For example, a TRE can comprise a tandem series of at least two, at least three, at least four, or at least five promoter fragments. Alternatively, a TRE can comprise one or more promoter regions along with one or more enhancer regions. TRE multimers can also comprise promoter and/or enhancer sequences from different genes. The promoter and enhancer components of a TRE can be in any orientation with respect to each other and can be in any orientation and/or any distance from the coding sequence of interest, as long as the desired cell-specific transcriptional activity is obtained.

The disclosed vectors are designed such that replication is preferentially enhanced in target cells in which the TRE(s) is (are) functional. More than one TRE can be present in a vector, as long as the TREs are functional in the same target cell.

A TRE for use in the present vectors may or may not comprise a silencer. The presence of a silencer (i.e., a negative regulatory element known in the art) can assist in shutting off transcription (and thus replication) in non-target cells. Thus, presence of a silencer can confer enhanced cell-specific vector replication by more effectively preventing replication in non-target cells. Alternatively, lack of a silencer may stimulate replication in target cells, thus conferring enhanced target cell-specificity.

As is readily appreciated by one skilled in the art, a TRE is a polynucleotide sequence, and, as such, can exhibit function over a variety of sequence permutations. Methods of nucleotide substitution, addition, and deletion are known in the art, and readily-available functional assays (such as the CAT or luciferase reporter gene assay) allow one of ordinary skill to determine whether a sequence variant exhibits requisite cell-specific

transcription regulatory function. Hence, functionally preserved variants of TREs, comprising nucleic acid substitutions, additions, and/or deletions, can be used in the vectors disclosed herein. Accordingly, variant TREs retain function in the target cell but need not exhibit maximal function. In fact, maximal transcriptional activation activity of a TRE may not 5 always be necessary to achieve a desired result, and the level of induction afforded by a fragment of a TRE may be sufficient for certain applications. For example, if used for treatment or palliation of prostate cancer, less-than-maximal responsiveness may be sufficient if, for example, the prostate cells are at an early stage of the disease.

10        Certain base modifications may result in enhanced expression levels and/or cell-specificity. For example, nucleic acid sequence deletions or additions within a TRE can move transcription regulatory protein binding sites closer or farther away from each other than they exist in their normal configuration, or rotate them so they are on opposite sides of the DNA helix, thereby altering spatial relationship among TRE-bound transcription factors, resulting 15 in a decrease or increase in transcription, as is known in the art. In one embodiment, the invention provides for certain modifications of a TRE that will result in modulated expression levels as directed by the TRE, including enhanced cell-specificity. Achievement of enhanced expression levels may be especially desirable in the case of more aggressive forms of neoplastic growth, and/or when a more rapid and/or aggressive pattern of cell killing is 20 warranted (for example, in an immunocompromised individual).

Transcriptional activity directed by a TRE (including both inhibition and enhancement) can be measured in a number of ways known in the art (and described in more detail below), but is generally measured by detection and/or quantitation of mRNA and/or of 25 a protein product encoded by the sequence under control of (i.e., operably linked to) a TRE.

As discussed herein, a TRE can be of varying lengths, and of varying sequence composition. The size of a heterologous TRE will be determined in part by the capacity of the viral vector, which in turn depends upon the contemplated form of the vector (see infra). 30 Generally minimal sizes are preferred for TREs, as this provides potential room for insertion of other sequences which may be desirable.

To minimize non-specific replication, endogenous (e.g., adenovirus) TREs are preferably removed from the vector. Besides facilitating target cell-specific replication,

removal of endogenous TREs also provides greater insert capacity in a vector, which may be of special concern if an adenoviral vector is to be packaged within a virus particle. Even more importantly, deletion of endogenous TREs prevents the possibility of a recombination event whereby a heterologous TRE is deleted and the endogenous TRE assumes transcriptional control of its respective adenovirus coding sequences (thus allowing non-specific replication). In one embodiment, an adenoviral vector is constructed such that the endogenous transcription control sequences of adenoviral genes are deleted and replaced by one or more heterologous TREs. However, endogenous TREs can be maintained in the adenovirus vector(s), provided that sufficient cell-specific replication preference is preserved.

5 These embodiments are constructed by inserting heterologous TREs between an endogenous TRE and a replication gene coding segment. Requisite cell-specific replication preference is determined by conducting assays that compare replication of the adenovirus vector in a cell which allows function of the heterologous TREs with replication in a cell which does not.

10

15 Generally, a TRE will increase replication of a vector in a target cell by at least about 2-fold, preferably at least about 5-fold, preferably at least about 10-fold more preferably at least about 20-fold, more preferably at least about 50-fold, more preferably at least about 100-fold, more preferably at least about 200-fold, even more preferably at least about 400- to about 500-fold, even more preferably at least about 1000-fold, compared to basal levels of 20 replication in the absence of a TRE. The acceptable differential can be determined empirically (by measurement of mRNA levels using, for example, RNA blot assays, RNase protection assays or other assays known in the art) and will depend upon the anticipated use of the vector and/or the desired result.

25 Replication-competent adenovirus vectors directed at prostate cells can be generated using TREs that are preferentially functional in a target cell. In one embodiment of the present invention, the prostate cell is a tumor cell. Non-limiting examples of tumor cell-specific heterologous TREs, and their respective target cells, include: probasin (PB), target cell prostate cancer (PCT/US98/04132). Methods for identification, isolation,

30 characterization and utilization of additional target cell-specific TREs are readily available to those of skill in the art.

In one embodiment of the present invention, a target cell-specific, heterologous TRE is tumor cell-specific. A vector can comprise a single tumor cell-specific TRE or multiple

heterologous TREs which are tumor cell-specific and functional in the same cell. In another embodiment, a vector comprises one or more heterologous TREs which are tumor cell-specific and additionally comprises one or more heterologous TREs which are tissue specific, whereby all TREs are functional in the same cell.

5

#### Prostate-specific TREs:

In one embodiment, adenovirus vectors comprise heterologous TREs that are prostate cell specific. For example, TREs that function preferentially in prostate cells and can be used to target adenovirus replication to prostate neoplasia, include, but are not limited to, TREs derived from the prostate-specific antigen gene (PSA-TRE) (Henderson U.S. Pat. No. 5,698,443); the glandular kallikrein-1 gene (from the human gene, hk2-TRE) (PCT US98/16312), and the probasin gene (PB-TRE) (PCT/US98/04132). All three of these genes are preferentially expressed in prostate cells and their expression is androgen-inducible. Generally, expression of genes responsive to androgen induction is mediated by an androgen receptor (AR).

15

#### Prostate-specific Antigen (PSA)

PSA is synthesized exclusively in prostatic epithelial cells and is synthesized in these cells whether they are normal, hyperplastic, or malignant. This tissue-specific expression of PSA has made it an excellent biomarker for benign prostatic hyperplasia (BPH) and prostatic carcinoma (CaP). Normal serum levels of PSA are typically below 5 ng/ml, with elevated levels indicative of BPH or CaP. Lundwall et al. (1987) FEBS Lett. 214:317; Lundwall (1989) Biochem. Biophys. Res. Comm. 161:1151; and Riegmann et al. (1991) Molec. Endocrin. 5:1921.

25

The region of the PSA gene that provides androgen-dependent cell specificity, particularly in prostate cells, involves approximately 6.0 kilobases (kb). Schuur et al. (1996) J. Biol. Chem. 271:7043-7051. An enhancer region of approximately 1.5 kb in humans is located between nt -5322 and nt -3739, relative to the transcription start site of the PSA gene. Within these enhancer sequences is an androgen response element (ARE) a sequence which binds androgen receptor. The sequence coordinates of the PSA promoter are from about nt -540 to nt +8 relative to the transcription start site. Juxtapositioning of the enhancer and promoter yields a fully functional, minimal prostate-specific TRE (PSA-TRE). Other portions

of this approximately 6.0 kb region of the PSA gene can be used in the vectors described herein, as long as requisite functionality is maintained.

Human Glandular Kallikrein (hk2)

5 Human glandular kallikrein (hk2, located on 19q13 and encoding the KLK2 protein) is expressed exclusively in the prostate and its expression is up-regulated by androgens, primarily through transcriptional activation. Wolf et al. (1992) Molec. Endocrinol. 6:753-762; Morris (1989) Clin. Exp. Pharm. Physiol. 16:345-351; Qui et al. (1990) J. Urol. 144:1550-1556; and Young et al. (1992) Biochem. 31:818-824. The levels of hK2 found in various 10 tumors and in the serum of patients with prostate cancer indicate that hK2 antigen may be a significant marker for prostate cancer. Charlesworth et al. (1997) Urology 49:487-493. Expression of hK2 has been detected in each of 257 radical prostatectomy specimens analyzed. Darson et al. (1997) Urology 49:857-862. The intensity and extent of hK2 expression, detected using specific antibodies, was observed to increase from benign 15 epithelium to high-grade prostatic intraepithelial neoplasia (PIN) and adenocarcinoma.

The activity of the hk2 promoter has been described and a region up to nt -2256 relative to the transcription start site was previously disclosed. Schedlich et al. (1987) DNA 6:429-437. The hk2 promoter is androgen responsive and, in plasmid constructs wherein the 20 promoter alone controls the expression of a reporter gene, expression of the reporter gene is increased approximately 10-fold in the presence of androgen. Murtha et al. (1993) Biochem. 32:6459-6464. hk2 enhancer activity is found within a polynucleotide sequence approximately nt -12,014 to nt -2257 relative to the start of transcription and, when this sequence is operably linked to an hk2 promoter and a reporter gene, transcription of 25 operably-linked sequences in prostate cells increases in the presence of androgen to levels approximately 30-fold to approximately 100-fold greater than the level of transcription in the absence of androgen. This induction is generally independent of the orientation and position of the enhancer sequences. Enhancer activity has also been demonstrated in the following regions (all relative to the transcription start site): about nt -3993 to about nt -3643, about nt -4814 to about nt -3643, about nt -5155 to about nt -3387, about nt -6038 to about nt -2394.

Thus, an hk2 enhancer can be operably linked to an hk2 promoter or a heterologous promoter to form a hk2 transcriptional regulatory element (hk2-TRE). A hk2-TRE can then

be operably linked to a heterologous polynucleotide to confer hk2-TRE-specific transcriptional regulation on the linked gene, thus increasing its expression.

Probasin

5       The rat probasin (PB) gene encodes an androgen and zinc-regulated protein first characterized in the dorsolateral prostate of the rat. Dodd et al. (1983) J. Biol. Chem. 258:10731-10737; Matusik et al. (1986) Biochem. Cell. Biol. 64:601-607; and Sweetland et al. (1988) Mol. Cell. Biochem. 84:3-15. The dorsolateral lobes of the murine prostate are considered the most homologous to the peripheral zone of the human prostate, where  
10      approximately 68% of human prostate cancers are thought to originate.

A PB-TRE has been shown to exist in an approximately 0.5 kb fragment of sequence upstream of the probasin coding sequence, from about nt -426 to about nt +28 relative to the transcription start site. This minimal promoter sequence from the PB gene appears to provide  
15      sufficient information to direct prostate-specific developmental--and hormone--regulated expression of an operably linked heterologous gene in transgenic mice. Greenberg et al. (1994) Mol. Endocrinol. 8:230-239.

Reporter Genes:

20       A target cell-specific TRE can be introduced into an adenovirus vector immediately upstream of and operably linked to an early gene such as E1A /AR chimera. The second gene may be a reporter gene. The reporter gene can encode a reporter protein, including, but not limited to, chloramphenicol acetyl transferase (CAT), beta-galactosidase (encoded by the lacZ gene), luciferase, alkaline phosphatase, a green fluorescent protein, and horse radish  
25      peroxidase. For detection of a putative cancer cell(s) in a biological sample, the biological sample may be treated with modified adenoviruses in which a reporter gene (e.g., luciferase) is under control of a target cell-specific TRE. The target cell-specific TRE will be transcriptionally active in cells that allow the target cell-specific TRE to function and luciferase will be produced. This production will allow detection of target cells, including  
30      cancer cells in, for example, a human host or a biological sample. Alternatively, an adenovirus can be constructed in which a gene encoding a product conditionally required for survival (e.g., an antibiotic resistance marker) is under transcriptional control of a target cell-specific TRE. When this adenovirus is introduced into a biological sample, the target cells

will become antibiotic resistant. An antibiotic can then be introduced into the medium to kill the non-cancerous cells.

**Preparation of the Adenovirus Vectors :**

5       The adenovirus vectors of this invention can be prepared using recombinant techniques that are standard in the art. Generally, a target cell-specific TRE is inserted 5' to the adenoviral gene of interest, preferably an adenoviral replication gene, more preferably one or more early replication genes (although late gene(s) can be used). A target cell-specific TRE can be prepared using oligonucleotide synthesis (if the sequence is known) or  
10      recombinant methods (such as PCR and/or restriction enzymes). Convenient restriction sites, either in the natural adeno-DNA sequence or introduced by methods such as PCR or site-directed mutagenesis, provide an insertion site for a target cell-specific TRE. Accordingly, convenient restriction sites for annealing (i.e., inserting) a target cell-specific TRE can be engineered onto the 5' and 3' ends of a UP-TRE using standard recombinant methods, such as  
15      PCR.

20       Polynucleotides used for making adenoviral vectors of this invention may be obtained using standard methods in the art, such as chemical synthesis, recombinant methods and/or obtained from biological sources.

Adenoviral vectors containing all replication-essential elements, with the desired elements (e.g., E1A) under control of a target cell-specific TRE, are conveniently prepared by homologous recombination or in vitro ligation of two plasmids, one providing the left-hand portion of adenovirus and the other plasmid providing the right-hand region, one or more of which contains at least one adenovirus gene under control of a target cell-specific TRE. If homologous recombination is used, the two plasmids should share at least about 500 bp of sequence overlap. Each plasmid, as desired, may be independently manipulated, followed by cotransfection in a competent host, providing complementing genes as appropriate, or the appropriate transcription factors for initiation of transcription from a target cell-specific TRE  
25      for propagation of the adenovirus. Plasmids are generally introduced into a suitable host cell such as 293 cells using appropriate means of transduction, such as cationic liposomes.  
30      Alternatively, in vitro ligation of the right and left-hand portions of the adenovirus genome can also be used to construct recombinant adenovirus derivative containing all the

replication-essential portions of adenovirus genome. Berkner et al. (1983) Nucleic Acid Research 11: 6003-6020; Bridge et al. (1989) J. Virol. 63: 631-638.

For convenience, plasmids are available that provide the necessary portions of adenovirus. Plasmid pXC.1 (McKinnon (1982) Gene 19:33-42) contains the wild-type left-hand end of Ad5. pBHG10 (Bett et al. (1994); Microbix Biosystems Inc., Toronto) provides the right-hand end of Ad5, with a deletion in E3. The deletion in E3 provides room in the virus to insert a 3 kb target cell-specific TRE without deleting the endogenous enhancer/promoter. The gene for E3 is located on the opposite strand from E4 (r-strand).  
5 PBHG11 provides an even larger E3 deletion (an additional 0.3 kb is deleted). Bett et al. (1994). Alternatively, the use of pBHGE3 (Microbix Biosystems, Inc.) provides the right hand end of Ad5, with a full-length of E3.  
10

For manipulation of the early genes, the transcription start site of Ad5 E1A is at 498 and the ATG start site of the E1A coding segment is at 560 in the virus genome. This region can be used for insertion of a target cell-specific TRE. A restriction site may be introduced by employing polymerase chain reaction (PCR), where the primer that is employed may be limited to the Ad5 genome, or may involve a portion of the plasmid carrying the Ad5 genomic DNA. For example, where pBR322 is used, the primers may use the EcoRI site in  
15 the pBR322 backbone and the XbaI site at nt 1339 of Ad5. By carrying out the PCR in two steps, where overlapping primers at the center of the region introduce a nucleotide sequence change resulting in a unique restriction site, one can provide for insertion of target cell-specific TRE at that site.  
20

A similar strategy may also be used for insertion of a target cell-specific TRE element to regulate E1B. The E1B promoter of Ad5 consists of a single high-affinity recognition site for Sp1 and a TATA box. This region extends from Ad5 nt 1636 to 1701. By insertion of a target cell-specific TRE in this region, one can provide for cell-specific transcription of the E1B gene. By employing the left-hand region modified with the cell-specific response  
25 element regulating E1A, as the template for introducing a target cell-specific TRE to regulate E1B, the resulting adenovirus vector will be dependent upon the cell-specific transcription factors for expression of both E1A and E1B. In some embodiments, part or all of the 19-kDa region of E1B is deleted.  
30

Adenoviral constructs containing an E3 region can be generated wherein homologous recombination between an E3-containing adenoviral plasmid, for example, BHGE3 (Microbix Biosystems Inc., Toronto) and a non-E3-containing adenoviral plasmid, is carried out.

5

Alternatively, an adenoviral vector comprising an E3 region can be introduced into cells, for example 293 cells, along with an adenoviral construct or an adenoviral plasmid construct, where they can undergo homologous recombination to yield adenovirus containing an E3 region. In this case, the E3-containing adenoviral vector and the adenoviral construct or 10 plasmid construct contain complementary regions of adenovirus, for example, one contains the left-hand and the other contains the right-hand region, with sufficient sequence overlap as to allow homologous recombination.

Alternatively, an E3-containing adenoviral vector of the invention can be constructed 15 using other conventional methods including standard recombinant methods (e.g., using restriction nucleases and/or PCR), chemical synthesis, or a combination of any of these. Further, deletions of portions of the E3 region can be created using standard techniques of molecular biology.

20

Methods of packaging polynucleotides into adenovirus particles are known in the art and are also described in PCT PCT/US98/04080.

25

The following examples are offered by way of illustration and should not be considered as limiting the scope of the invention. The specific examples exemplify the adenovirus 5 serotype, however, persons skilled in the art will realize these techniques may be applied to other adenoviral serotypes

#### Delivery of Adenoviruses:

Delivery of adenoviral vectors is discussed infra and is generally accomplished by 30 either site-specific injection or intravenously. Direct intra-prostatic injections are preferred. Site-specific injections of either vector may include, for example, injections into the portal vein of the liver as well as intraperitoneal, intrapleural, intrathecal, intra-arterial, intra-tumor injections or topical application. These methods are easily accommodated in treatments using adenoviral vectors.

The adenoviral vectors may be delivered to the target cell in a variety of ways, including, but not limited to, liposomes, general transfection methods that are well known in the art (such as calcium phosphate precipitation or electroporation), direct injection, and intravenous infusion. The means of delivery will depend in large part on the particular 5 adenoviral vector (including its form) as well as the type and location of the target cells (i.e., whether the cells are to be transfected or transformed *in vitro* or *in vivo*). If used as a packaged adenovirus, adenovirus vectors may be administered in an appropriate physiologically acceptable carrier at a dose of about 1 to about 10. The multiplicity of infection will generally be in the range of about 0.001 to 100. If administered as a 10 polynucleotide construct (i.e., not packaged as a virus) about 0.01 ug to about 1000 ug of an adenoviral vector can be administered. The adenoviral vector(s) may be administered one or more times, depending upon the intended use and the immune response potential of the host, and may also be administered as multiple, simultaneous injections. If an immune response is undesirable, the immune response may be diminished by employing a variety of 15 immunosuppressants, so as to permit repetitive administration, without a strong immune response. If packaged as another viral form, such as HSV, an amount to be administered is based on standard knowledge about that particular virus (which is readily obtainable from, for example, published literature) and can be determined empirically.

20 Administration of the above-described methods may also include repeat doses or courses of target-cell specific adenovirus depending, *inter alia*, upon the individual's response and the characteristics of the individual's disease. Repeat doses may be undertaken immediately following the first course of treatment (i.e., within one day), or after an interval of days, weeks or months to achieve and/or maintain suppression of tumor growth. A 25 particular course of treatment according to the above-described methods, for example, adenoviral therapy, may later be followed by a course of combined or separate radiation and adenoviral therapy.

Generally, an effective amount of adenovirus vector is administered, i.e., amounts 30 sufficient to achieve the desired result, based on general empirical knowledge of a population's response to such amounts. Some individuals are refractory to these treatments, and it is understood that the methods encompass administration to these individuals. The amount to be given depends, *inter alia*, on the stage of prostate cancer, the condition of the

individual, the extent of disease, the route of administration, how many doses will be administered, and the desired objective.

**Methods of Treating Prostate Cancer:**

5 One aspect of the invention pertains to a method for treating prostate cancer in a subject in need of such treatment, comprising administering to the subject a adenoviral particle of the invention, and performing on the subject at least one procedure that removes or destroys prostatic tumor tissue, such as a radical prostatectomy, cryosurgery, external radiation therapy (e.g., X-ray therapy) or interstitial radiation therapy (e.g., implantation of a  
10 radioactive seed). The adenoviral particle may be administered to the subject prior to or subsequent to performing the procedure that removes or destroys prostatic tumor tissue. In one such embodiment, administration of an adenoviral particle is preferably for a period sufficient to cause the prostate or prostatic tumor tissue to shrink in size prior to performing the procedure that removes or destroys prostatic tumor tissue. A suitable period for  
15 preadministration of an adenoviral particle typically is between about one day and about one year, more preferably between about three days and about six months.

In certain situations it may be desirable to use an antiandrogen, and thus in another embodiment, this treatment method can further involve administering an antiandrogen to the  
20 subject in combination with the adenoviral particle. In yet another embodiment, this treatment method can further involve administering one or more inhibitors of sex steroid biosynthesis to the subject in combination with the peptide compound (optionally in further combination with an antiandrogen) prior to or subsequent to performing the procedure that removes or destroys prostatic tumor tissue.

25 In another embodiment, the adenoviral particle of the present invention may be administered in conjunction with an LHRH agonist, as described in U.S. Patent Nos. 5,843,902, 5,780,435, and 6,153,586, incorporated herein by reference, or an LH receptor antagonist.

30 Those of skill in the art will recognize that while it may not be necessary to combine adenoviral particle therapy with additional drugs or treatments, in certain situation it may be desirable to further combine the compound with other drugs or treatments to achieve the greatest therapeutic effect.

The methods of the present invention can be applied to the treatment of prostate cancer in male subjects at any stage of the cancer, although certain treatment methods are more preferred for particular cancer stages. For reviews on screening and diagnostic methods for prostate cancer, see e.g., Garnick, M. (1993) Annals of Internal Medicine 118:803-818; 5 and Garnick, M. (1994) Scientific American 270:72-81. Prostate cancer is commonly evaluated according to a scale divided into four lettered stages: A, B, C and D. Tumors in stage A are microscopic; stage A<sub>1</sub> designates tumors confined to a relatively small area and composed of well-differentiated tissue, while stage A<sub>2</sub> tumors are more diffuse and less well differentiated. Stage B tumors are large enough to be felt during a rectal examination, while 10 stage C prostate cancers have spread throughout the gland and typically have pushed past the borders of the prostate into surrounding structures. Stage D tumors have metastasized, e.g., to lymph nodes, bone, or other organs. Alternatively, tumors can be staged by the TNM staging system, in which tumors are ranked on a scale of progressively worsening disease from T1a to T4 (e.g., T1c tumors are non-palpable and non-visible that were detected by elevated blood 15 levels of prostate specific antigen). The methods of the invention are useful in the treatment of any stage of prostate cancer. However, it will be appreciated by the skilled artisan that methods involving procedures for removal or destruction of prostatic tumor tissue preferably are employed with non-metastasized cancers. For example, radical prostatectomy preferably is used with stage A, B and some stage C tumors (i.e., where the tumor growth has not 20 extended considerably beyond the borders of the prostate gland) as well as stage T1c tumors. Radiation therapy (e.g., external or interstitial) preferably is used with stage A, B or C tumors as well as T1c tumors.

To assess the efficacy of a treatment method of the invention, the size of the prostate 25 can be determined by methods known in the art, for example, rectal examination, transrectal ultrasonography or magnetic resonance imaging (MRI). Moreover, the size or extent of the prostate tumor (and metastatic tumors, if any) can be assessed by known methods including a prostate-specific antigen blood test (described further below), bone scanning, X-rays, skeletal survey, intravenous pyelography, CAT-scan, MRI, physical examination, biopsy, and the 30 like. For treatment methods that involve surgery (e.g., in neoadjuvant therapy wherein an adenoviral particle is administered prior to a radical prostatectomy), the tumor can also be staged during surgery (e.g., the prostate gland can be examined during surgery and/or a biopsy can be taken and examined). Thus, clinical staging and/or surgical staging may be used to evaluate the extent of disease. Use of an adenoviral particle in accordance with the

methods of the invention is expected to result in a tumor stage, which is improved compared to methodologies that do not include use of an adenoviral particle of the invention.

A preferred method of evaluating the extent of prostate cancer is to assay the level of prostate-specific antigen (PSA) in a subject's blood. The PSA blood test is a reasonably specific, sensitive, rapid, and inexpensive tool for screening for prostate cancer. In general, a blood PSA level above 4 ng/ml is considered to be suggestive of the presence of prostate cancer, with levels above 10 ng/ml being particularly indicative of cancer. For a subject undergoing treatment with an adenoviral particle, according to the methods of the invention, a pretreatment level of PSA can be established and the efficacy of the treatment assessed by monitoring periodically the PSA level in the subject's blood, wherein decreased PSA levels are used as an indicator of the efficacy of the treatment. The PSA nadir (i.e., the point at which PSA levels do not decrease further even upon further treatment with a peptide compound) can be used as the indicator point for initiation of a second therapy, for example for performance of a procedure that removes or destroys prostatic tumor tissue (such as radical prostatectomy, cryosurgery and/or radiation therapy). It is expected that the PSA nadir will be reached sooner using an adenoviral treatment, as compared to treatments which do not include using an adenoviral treatment.

The response criteria for prostate developed by the National Prostate Cancer Project (see e.g., *The Prostate*, 1:375-382) can also be used to assess the efficacy of treatment. For treatment methods involving a procedure that removes or destroys tumor tissue (such as radical prostatectomy, cryosurgery, and/or radiation therapy), it is preferable to administer oncolytic adenoviral therapy until the size of the prostate or a prostate tumor has decreased and/or blood PSA levels have decreased before performing the procedure.

### **Cessation of Viral Replication**

Conditionally replication competent adenoviruses currently do not have any active means of inhibiting the replicative cycle, other than those mechanisms utilized by the host. While the host immune system is capable of reversing such infections, some patients may have varying degrees of immunocompetence. In certain situations, it may be possible that an infection proceeds in a fashion which is detrimental to the patient. In such a circumstance, having the ability to immediately halt an active replicative cycle would be highly desirable. The androgen receptor, as described above, is regulated in part by its associations with certain

chaperonins, including hsp90. Certain compounds, including the ansamycins (of which geldanamycin is prototypical), are capable of inhibiting hsp90 and result in the immediate inactivation of those proteins which are found to be complexed to that chaperonin. The androgen receptor is such a protein, as is the E1A-AR chimeras described in this invention.

5     Addition of geldanamycin results in the immediate inactivation of the E1A-AR chimeras, and immediately halts viral replication of those constructs based on these chimeras. Since at least two ansamycins (geldanamycin and 17-AAG also known as 17-allylamino-17-demethoxygeldanamycin) are currently in clinical trials, this strategy provides a novel "off-switch" for tissue specific replication competent adenoviral therapy.

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#### **Other Potential Uses of E1A Nuclear Receptor Chimeras**

Although the methods of the invention are described in particular with application to the treatment of prostate cancer, it will be appreciated by the skilled artisan that these methods also can be applied to the treatment of other disorders, including other cancers. For 15 example, a chimeric fusion of the E1A gene with other nuclear receptors such as the estrogen receptor, progesterone receptor, thyroid hormone receptor or retinoic acid receptor, may allow differential regulation of certain promoters which would markedly enhance tissue specific replication in a ligand dependent fashion. Such an approach could prove highly efficacious for breast cancer, thyroid cancer or any other cancer which relies upon a particular 20 nuclear receptor for part of its development or propagation. The paradigm would be similar to the one described for the androgen receptor and its relationship to prostate cancer.

This invention is further illustrated by the following examples which should not be construed as limiting. The contents of all references, patents, and published patent 25 applications cited throughout this application, as well as the figures and the sequence listing, are incorporated herein by reference.

#### **EXAMPLES**

#### **30 MATERIALS AND METHODS**

Unless indicated otherwise, the following materials and methods were used in the subsequent examples.

**Cells and Cell Culture.** LNCaP, LAPC4 and PC3, DU145 are human prostate cancer cell lines. Hela 3S is a human cervical cancer cell line. All were obtained from ATCC (Manassas, VA). LNCaP, PC3 and DU145 were maintained in RPMI medium supplemented with 10% FBS. Hela 3S and 293 cells (Quantum Biotech) were maintained in Dulbecco's MEM with 10% FBS. LAPC4 was maintained in ISCOVE's medium with 10%FBS and 1nM R1881. All cells were maintained at 37°C in an atmosphere containing 5% CO<sub>2</sub>.

**Plasmid Construction.** E1A fragment was synthesized (approximately 1kb) from the wild type adenovirus type 5 by PCR using the primer pair of 5'-

10 TCACTCGGATCCACCGGGACTGAAAATGAGACATAT and 5'-TACATCACTC  
GCAGGCCGCTGGCCTGGGGCGTTACAGCTCA. This E1A product was cloned into the  
vector pCR2.1-TOPO (Invitrogen) to generate TOPO-E1A plasmid, in which the E1A  
sequence was verified by sequencing analysis. Full-length of androgen receptor (AR full-  
length), AR without ligand-binding domain (AR ORF 1-1959 bps, named AR TAD-DBD),  
15 and AR DNA-binding domain alone (AR ORF 1669-1959 bps, named AR DBD) were  
synthesized from the AR cDNA by PCR using the primer pairs of 5'  
TACATCACTCGCGGCCGAGAAGTCAGTTAGGGCTGGAA and 5'-TCAC  
TCCTCGAGTC ACTGGGTGTGGAAATAGATGGC TT for AR full-length (approx. 2.7  
kb), 5'-TAC ATCACTCGCGGCCGAGAAGTCAGTTAGGGCTGGG AA and 5'-  
20 TCACTCCTC GAGTCACTCAGTGGGCTGGTGGT GCTGGA for AR-TAD-DBD  
(approx. 2 kb), 5'-TACATCACTCGCGGCCGAAAGACCTGCCTGATCTGTGGAGAT  
and 5'-TCACTCCTCGAGTCACTCAGTGGGCT GGTGGTGCTGGA for AR-DBD  
(approx. 320 bps). These three different lengths of AR PCR products were cloned into the  
vector pCR2.1-TOPO to generate the plasmids of TOPO-AR full-length, TOPO-AR TAD-  
DBD, TOPO-AR DBD, in which the sequence of these AR inserts were verified by  
25 sequencing analysis. The AR fragments were cleaved from TOPO-AR by Not I and Xho I  
restriction enzyme digestion and inserted at the downstream of the TOPO-E1A to create  
TOPO-E1A-AR. The E1A-AR fragments were cleaved from TOPO-E1A-AR plasmids by  
BamH I and Xho I and cloned into pBK-CMV to create the pBK-CMV/E1A-AR constructs.  
30 The E1A fragment was cloned into pBK-CMV to create the pBK-CMV/E1A construct. For  
cloning of E1A reporter plasmid pE1B-luc, E1B promoter with Hind III and BamH I ends  
was synthesized from Ad5 wild type virus by PCR amplification using primer pair 5'-  
CCCCAGCTTCCTCTAACACACACCTCCTG and 5'-CGGGATCCGA  
GGTCAGATGTAACCAAGA. The Luciferase gene was cleaved from the plasmid pBK-

PSE-PBN-Luc with BamHI and XmaI. The PCR product of E1B promoter and the Gel purified luciferase were cloned into the pUC19 vector with ends of Hind III and Xma I, generating the E1A reporter plasmid pE1B-luc. The plasmid pcDNA3-hAR was a kindly gift from Dr. William Isaacs. To construct the plasmids that the E1A-AR gene expressions are 5 under the control by the prostate specific enhancer (PSE) and rat probasin promoter (PBN), the prostate specific reporter plasmid pBK-PSE-PBN-luc was digested with BamHI and XhoI to remove the luciferase gene. The large fragments containing pBK-PSE-PBN were gel-purified and ligated with the BglII-XhoI fragments of E1A-AR inserts cleaved from pBK-CMV/E1A-AR, generating the plasmids pBK-PSE-PBN-E1A-AR. The E1A-AR chimera in 10 the pBK-PSE-PBN-E1A-AR plasmids includes wild type AR (E1A-AR), transactivation domain of AR (E1A-TAD), and a full-length AR with a point mutation C685Y in the ligand binding domain (E1A-AR<sup>C685Y</sup>). The plasmid pBK-PSE-PBN-E1A was constructed by replacing the luciferase gene in the plasmid pBK-PSE-PBN-luc with E1A fragment. All 15 PCR-derived fragments and site-directed mutagenesis were sequenced to confirm their predicted composition.

**Construction of shuttle vectors for homologous recombination in bacteria.** To generate our recombinant constructs, the original pShuttle vector from AdEasy System was modified as following: an internal BamH I site was ablated and re-introduced into multiple cloning sites (MCS) for insertion of our favorite genes when combined with other restriction enzymes. Our recombinant adenoviruses were designed to have an intact E1B, however, the adenoviral backbone vectors pAdEasy-1 does not contain Ad5 sequences encompassing the E1 genes (nucleotides 1–3,533) and the original pShuttle has impaired E1B. We re-introduced an intact E1B by double digestion of the adenoviral plasmid pFG173 (purchased 20 from Microbix Biosystems) with Hpa I and Bst1107I, this 4193 nucleotide fragment corresponding to the right hand portion of the adenovirus from the E1A polyadenylation signal through into E1 replaced the 2362 nucleotide fragment in the original pShuttle vector, the subsequent plasmid named RpS-TOAD-shuttle (refer to Tissue specific Oncolytic Ad Shuttle vector). To generate shuttle plasmids RpS-PSE-PBN-E1A-AR (AR wt named RP103, 25 AR TAD named RP104, AR<sup>C685Y</sup> named RP105), the PSE-PBN-E1A-AR fragments were cleaved from plasmid pBK-PSE-PBN-E1A-AR at Bgl II and Xho I sites and then ligated with the shuttle vector RpS-TOAD that was linearized with BamH I and Xho I. The positive control shuttle plasmid RpS-PSE-PBN-E1A (RP101) was generated by insertion of PSE- 30 PBN-E1A fragment into the RpS-TOAD at BamH I and Not I sites. The shuttle plasmid RpS-

PSE-PBN-luc (RP106) was generated by insertion of PSE-PBN-luc fragment into the RpS-TOAD at BamH I and NotI sites. The negative control shuttle plasmid RpS-PSE-PBN-null (RP102) was generated by insertion of PSE-PBN-null fragment into the RpS-TOAD at BamH I and Sal I sites.

5

**Generation of Recombinant Adenoviruses.** The AdEasier-1 cells were used for transformation of all shuttle plasmids. AdEasier-1 cells are high-recombinant *E. coli* strain BJ5183 derivatives that have stable transfection of adenoviral backbone AdEasy-1 genome (E1 and E3 region were deleted) to allow for rapid homologous recombination with single step transformation and selection. The shuttle plasmids were linearized with PmeI and then transformed into the AdEasier-1 cells. The clones that had inserts were further characterized with extensive restriction digestion. The digestion patterns were compared with the pAdEasy-1 stock DNA made in a non-recombinant strain. The right clones after screening were transformed into non-recombinant strain DH10B cells for large-scale amplification. 10 These recombinant plasmids were linearized with PacI and transfected into adenovirus packaging cell line 293 cells to generate our recombinant adenoviruses. 15

**Transfection and Luciferase Assay.** Cells were plated at  $1 \times 10^4$  per well (triplicates) in 96-well plate at 1 day prior to the transfection in antibiotic-free medium with 10% FBS. 20 100 ng of the reporter plasmid pBK-PSE-PBN-luc (*firefly* luciferase gene under control of the prostate specific enhancer and promoter) and 1 ng of the reporter plasmid pRL-luc (*Renilla* luciferase gene under control of the CMV promoter) were co-transfected with 100 ng of the other interest plasmids using Lipofectamine Plus® Reagent (Gibco). Cells were added OPTI-MEM® medium (Gibco) with dextran-charcoal-stripped serum (10% final concentration) in 25 the presence or absence of synthetic androgen R1881 (Methyltrienolone, Sigma) (final concentration 5nM) at 3 hours post-transfection. The Luciferase assays (Dual-Glo Luciferase Assay System, Promega) were performed at 24-48 hours post-transfection. All of the transfection assays were triplicates and normalized to the reporter plasmid pRL-luc.

30 **Example 1 E1A Inhibits AR Activation of a Reporter Gene:**

Since the androgen receptor and E1A share many common interactions with mediators of target gene expression, we hypothesized that E1A expression may adversely affect AR activity. In order to test this hypothesis, we cloned the E1A gene from the adenovirus wildtype genome (including both introns 1 & 2) into an expression plasmid (pBK-CMV from

Stratagene), under the constitutive control of the CMV promoter. We then performed co-transfection experiments with an androgen inducible promoter (pBK-PSE-PBN-LUC), described above. We chose to use LAPC4 as the AR positive cell line for our analysis, since this cell line expresses wildtype AR, while the more commonly used LNCaP expresses a mutant AR; however, to date we have consistently found similar results with both cell lines when performing androgen induction. PC3 was used as the prototypical AR negative prostate cancer cell line. Figure 1, Panel A demonstrates the results of co-transfection of the E1A construct and the androgen inducible promoter PSE-PBN-LUC in LAPC4. In the first lane the reporter (pBK-PSE-PBN-LUC) was co-transfected with vector only (pBK-CMV, no insert) in the absence of hormone (R1881). In the next lane, the same conditions were used, but in the presence of 10 nM R1881 (a synthetic androgen). This demonstrated the expected induction of the target reporter gene by the androgen receptor. Incubation with plasmid encoding the E1A gene, however, reversed the hormone induction of the target gene expression. These results were performed over a wide range of plasmid concentrations (.1 µg to 1 µg) and demonstrated a clear dose-response relationship between E1A expression and inhibition of AR target gene induction; though at concentrations of the E1A plasmid > 0.5 µg inhibition of basal expression (i.e., not AR dependent) could also be achieved. The next issue was whether or not the amount of E1A expression required for this inhibition was relevant compared to the amount expressed in typical viral infections. Thus we also performed the same transfection experiment in LAPC4, followed however by infection with either a control virus (PSE-NULL, which is replication deficient and has no E1A trans-gene insert) or an E3 deleted wildtype replicating virus (CN702). Because these viruses can cause replication in LAPC4, we assayed the effect on target gene expression at 24 hours post-infection to minimize any bias-effect from the lytic cycle on our target gene expression. These data show that while the control virus PSE-Null has roughly the same induction of target gene expression, as the target plasmid alone in the presence of R1881, the wt-E3 deleted CN702 clearly demonstrated a dose response inhibition of AR activity, consistent with our transfection data. By contrast, repeating the exact same experiments in a non-AR expressing prostate cancer cell line (Figure 1, Panel B, PC3) demonstrated no appreciable induction of target gene expression with androgen. Moreover, in PC3, E1A expression by plasmid or by virus showed only a nominal decrease in the target gene expression---a result also seen with our replication deficient PSE-Null virus, indicating that this is a nonspecific finding. We conclude from these studies that E1A causes direct inhibition of AR function. Similar

experiments in LNCaP reveal that the same inhibition occurs (data not shown). In fact, high levels of E1A are capable of suppressing many tissue specific promoters, even in the absence of AREs or AR. An example of this is shown in Figure 2. In this experiment the probasin promoter, the PSA promoter and the PSMA promoter were all tested, with and without AREs 5 (supplied by fusion of the promoter with the PSA enhancer, aka "PSE") in both LAPC4 and PC3 cells again. Unlike before, here we over-expressed E1A by increasing the pBK-CMV-E1A expression plasmid by 5 fold. All the promoters shown were tested in the presence of 10 nM R1881. The addition of PSEs to the basal promoters did result in an enhancement of androgen dependent transactivation of the reporter genes in LAPC4, but not PC3, as 10 expected. However, over-expression of E1A results in inhibition of transgene expression even in the AR negative PC3 cell line. These data indicate that E1A has a tendency to inactivate *prostate specific genes* in general and thus enhancement of prostate specific oncolytic gene therapy will require methods that overcome this inhibition.

15 As outlined above, there are at least two possible mechanisms by which E1A could result in inhibition of AR function: (1) sequestration of common transcriptional regulatory complexes (2) direct interaction with CtBP in an inhibitory complex. In order to help us resolve these two possibilities, we attempted co-immunoprecipitation experiments with antibodies directed against E1A and AR. Despite multiple attempts, we were never able to 20 demonstrate direct interactions between the two proteins (data not shown). Therefore, we conclude that the most probable mechanism was sequestration of the shared transcriptional regulatory factors between E1A and AR. If such a mechanism is correct, we predicted that fusion of the androgen receptor with the E1A gene would convert E1A from a repressor of AR function to a potent co-activator. Thus, we generated several E1A-AR fusion constructs. 25 The major constructs included E1A-AR, E1A-AR-C685Y, E1A-TAD, and E1A-DBD. The AR corresponds to the 2.7 Kb full-length cDNA, fused in frame with E1A by PCR cloning. TAD stands for transactivation domain (amino terminus including AF-1, DNA binding domain and the hinge region). DBD corresponds to the DNA binding domain (both zinc fingers and hinge region). E1A-AR-C685Y corresponds to a fusion containing the full length 30 AR cDNA with a single mutation at codon 685 converting cysteine to tyrosine. This point mutation results in an androgen receptor which is fully activated with nonsteroidal androgen receptor antagonists, such as flutamide and bicalutamide

**Example 2: AR-E1A Fusion Chimera Markedly Enhances AR dependent Trans-gene Activation:**

We tested our hypothesis that a fusion of E1A and AR domains would convert E1A into a potent co-activator by performing co-transfection of our fusion constructs with an AR dependent reporter construct pBK-PSE-PBN-LUC (Figure 3). Each well of a 96 well plate was co-transfected with 0.1 µg of an E1A fusion construct, and a fixed amount of pBK-PSE-PBN-LUC (0.1 µg) in the presence or absence of androgen (R1881) in three different cell lines. The first two cell lines were prostate cancer; the last was non-prostate (HeLa). The AR positive line LNCaP demonstrated induction of the reporter with androgen (control), inhibition of induction with E1A, and constitutive expression with E1A-TAD. The full-length AR fusion (E1A-AR) demonstrated an excellent activation with hormone (super-induction), both in the AR positive cell line LNCaP and the AR negative cell line PC3. The E1A-DBD raised basal expression of the reporter slightly in all cell lines in a non-specific and hormone independent fashion. Importantly, the HeLa line failed to demonstrate much activity of any of the constructs with or without androgen. Subsequent experiments confirm the same pattern in LAPC4 as LNCaP and lack of activity in the colon cancer cell line DLD. Thus, the fusion of the E1A-AR is able to convert the AR negative cell line PC3 into an active inducer of prostate specific promoter activity, while maintaining specificity of action. This finding has tremendous implications as it suggests that an oncolytic virus based on such a construct could be useful in patients who have a subpopulation of prostate cancer cells which have lost expression of the androgen receptor. Moreover, it recently occurred to us that a simple point mutation in the AR ligand-binding domain (C685Y) could convert the process from an androgen dependent activation, to an anti-androgen dependent activation. Therefore we recently generated the E1A-AR-C685Y construct by site directed mutagenesis. Our preliminary experiments reveal potent stimulation by bicalutamide to stimulate expression of the AR dependent reporter gene pBK-PSE-PBN-LUC and significant activation by R1881, as expected.

**Example 3. The Effect of Geldenamycin on Androgen Receptor function in EA1/Ar chimera transfected PC-3 cells and COS-1 cells:**

As illustrated in Figures 20 and 21, different chimeras were tested in two different cell lines (PC3 and COS-1 cells) in the presence of hormone (R1881) with and without. Geldenamycin. The results indicate that Geldenamycin can inactivate the effect on expression

by the three chimeras, E1A/AR (full length), E1A/TAD and E1A/C685Y. RLU are expressed as % Renilla control co-transfection expression (i.e., dual luciferase).

**Equivalents**

5        Those skilled in the art will recognize, or be able to ascertain using no more than routine experimentation, many equivalents to the specific embodiments of the invention described herein. Such equivalents are intended to be encompassed by the following claims.